

MRC

Brain Network  
Dynamics Unit



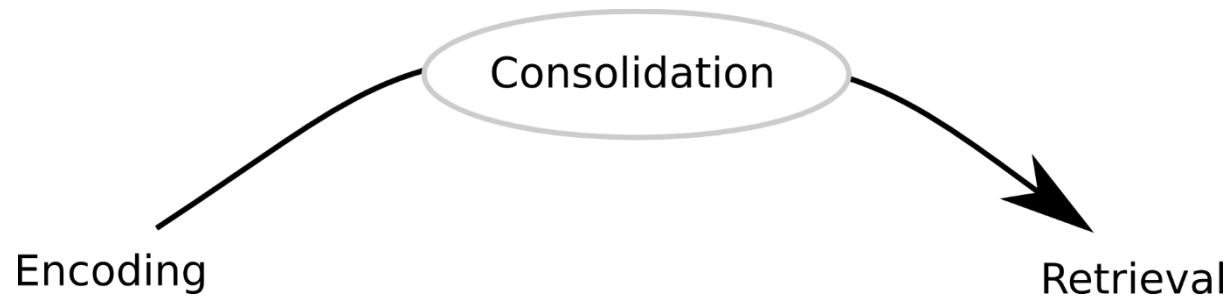
UNIVERSITY OF  
**OXFORD**

# Hippocampal reactivation stabilizes recently formed cell assembly patterns

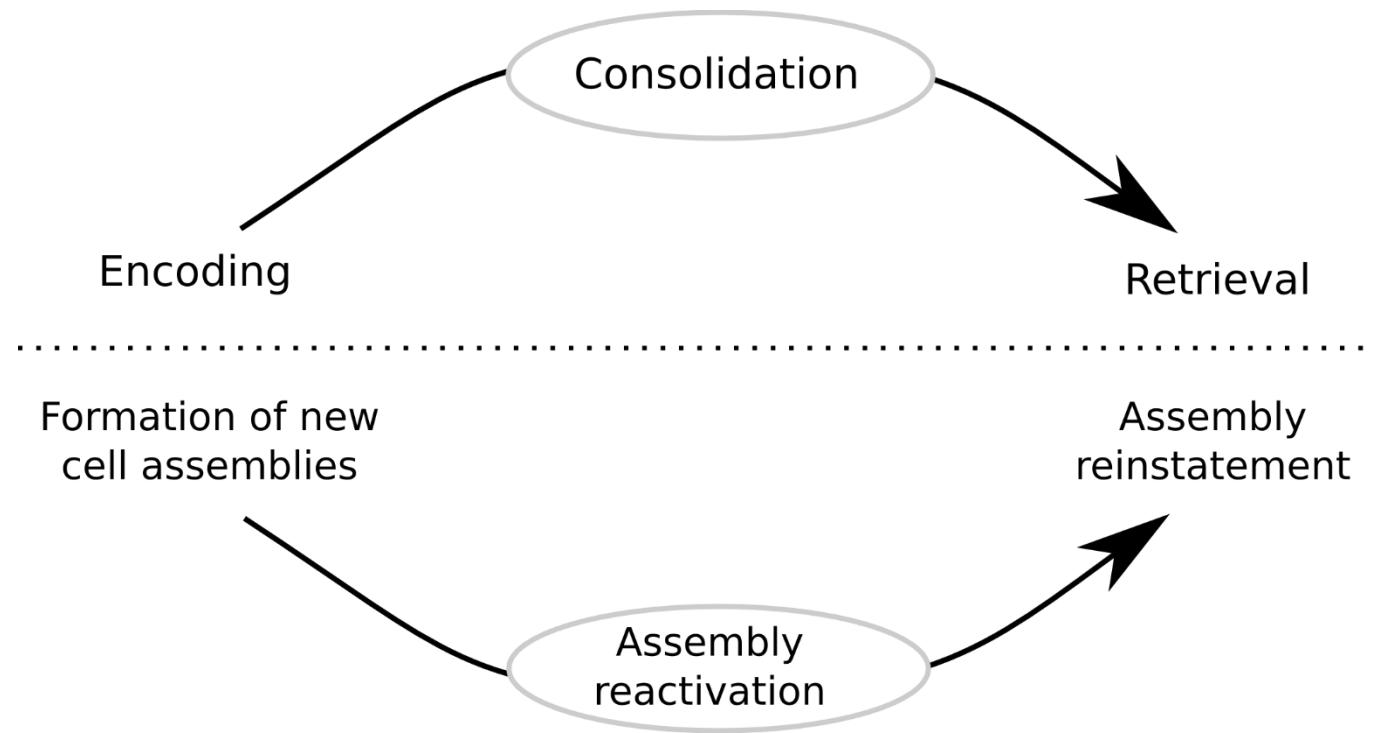
*Gido van de Ven*

8 May 2017, Cardiff

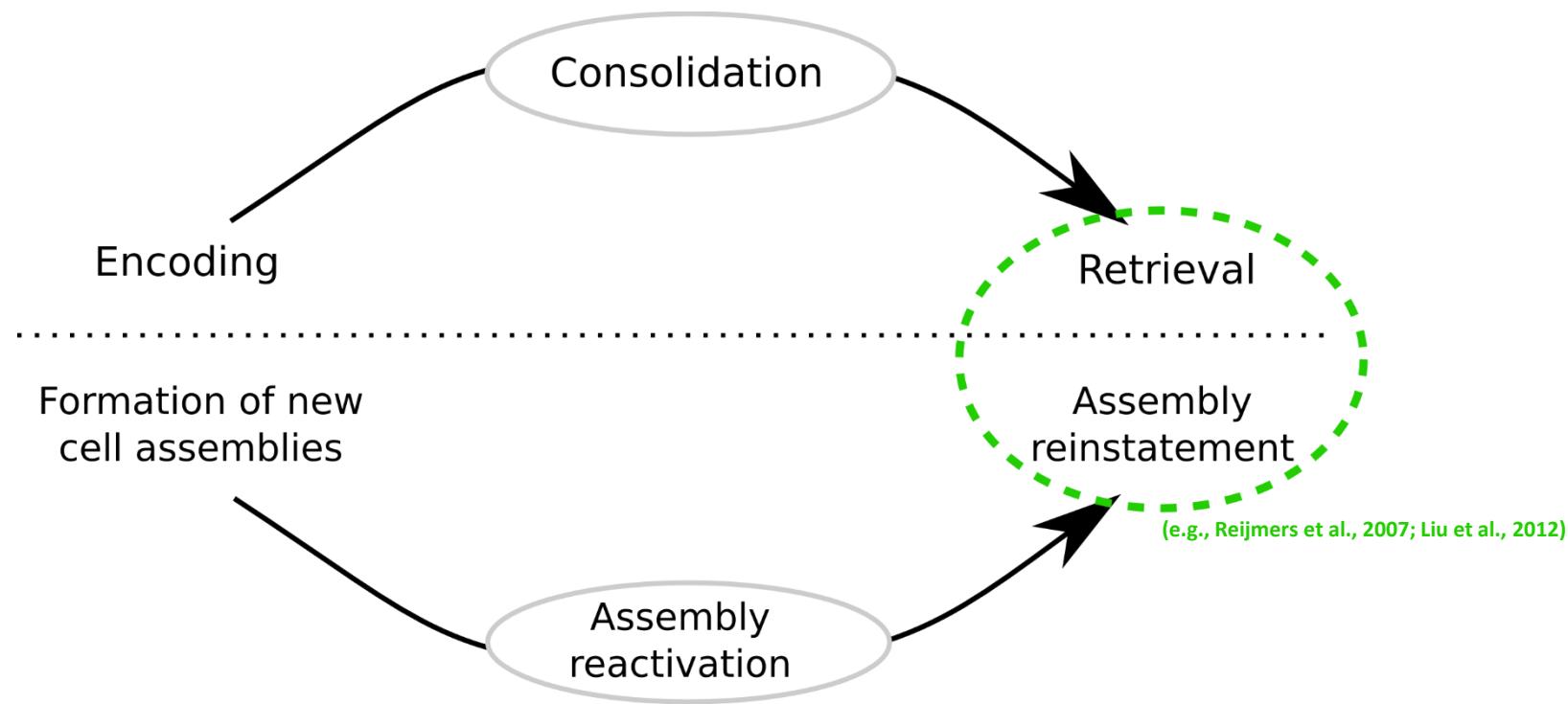
# Memory-consolidation



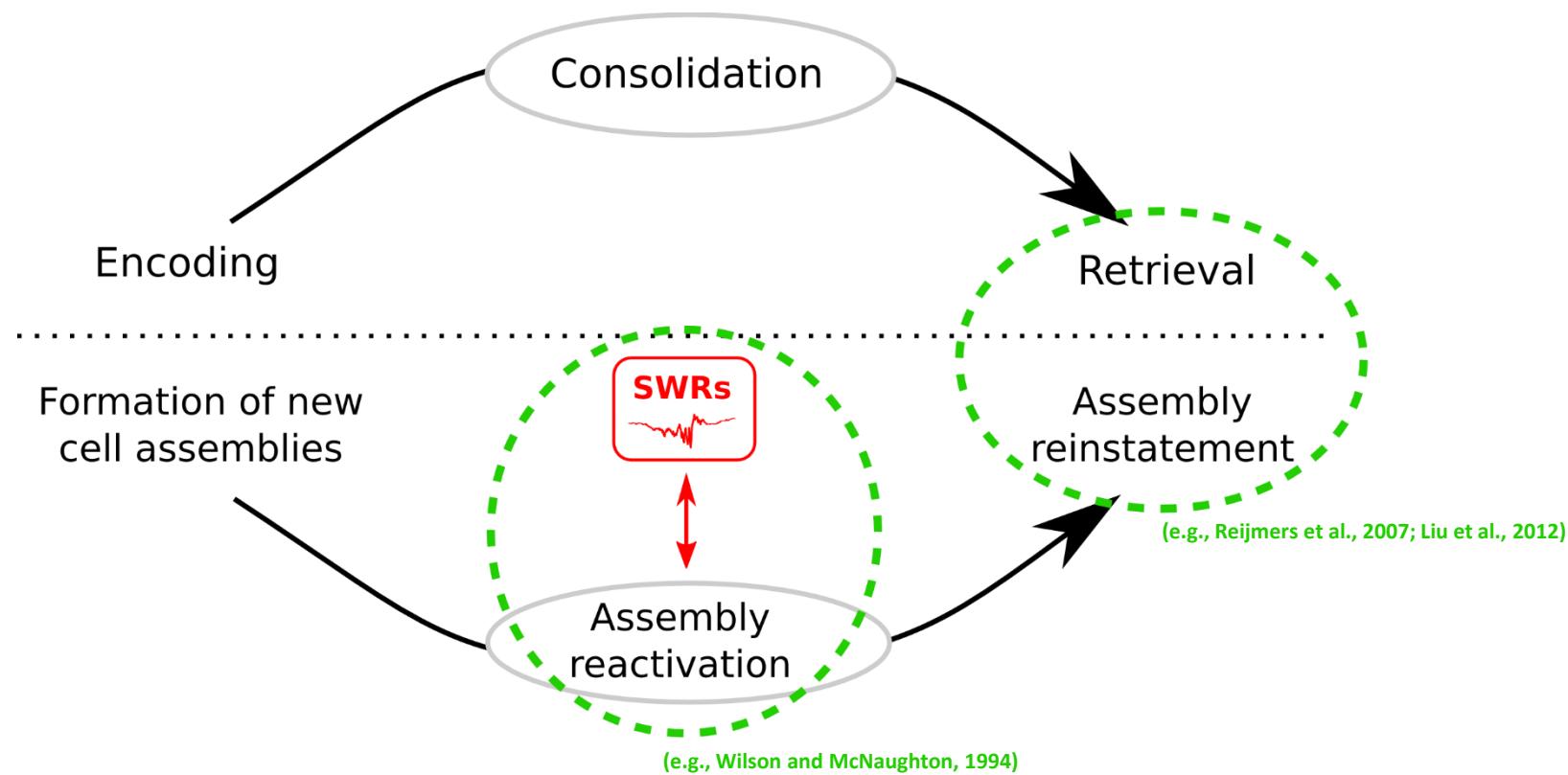
# Memory-consolidation: cell assembly / reactivation hypothesis



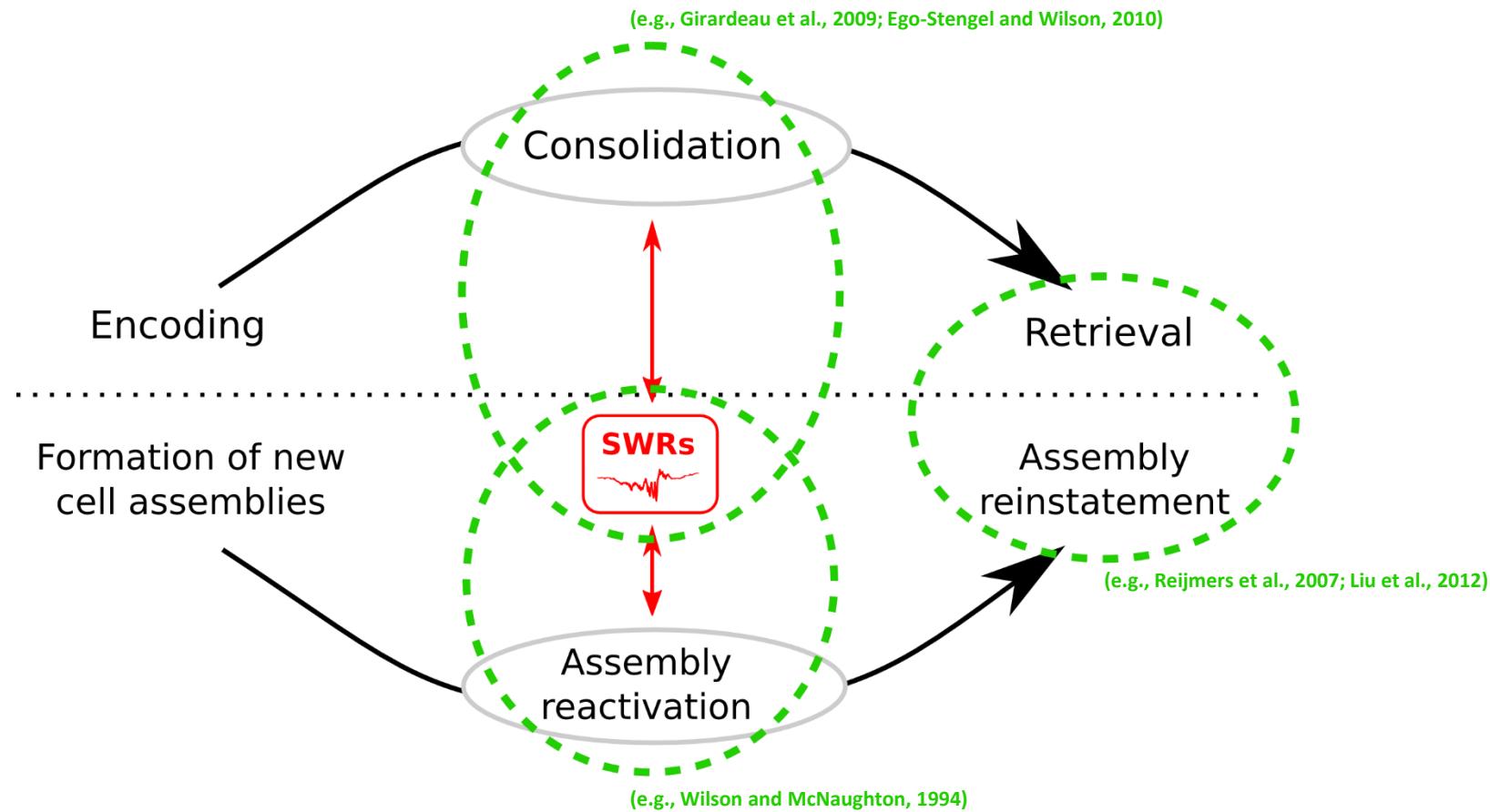
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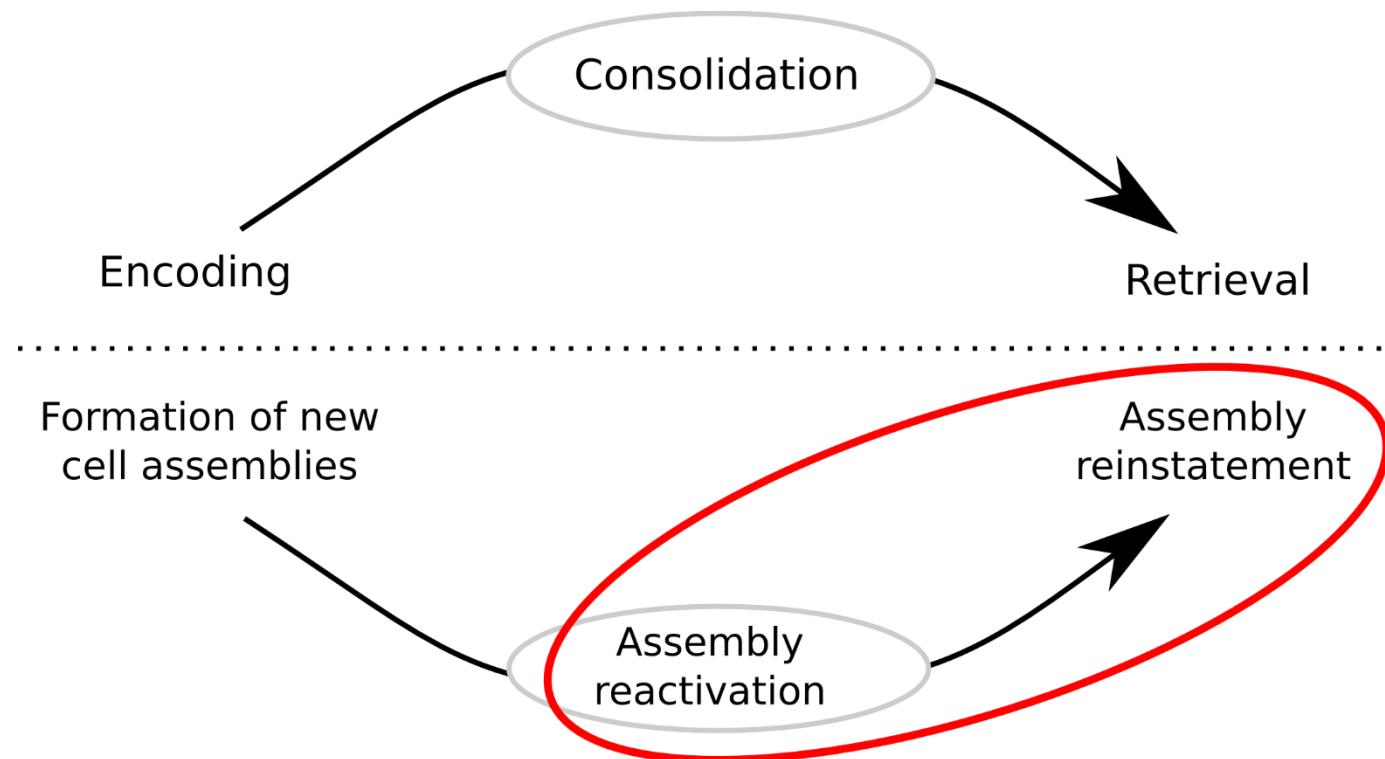
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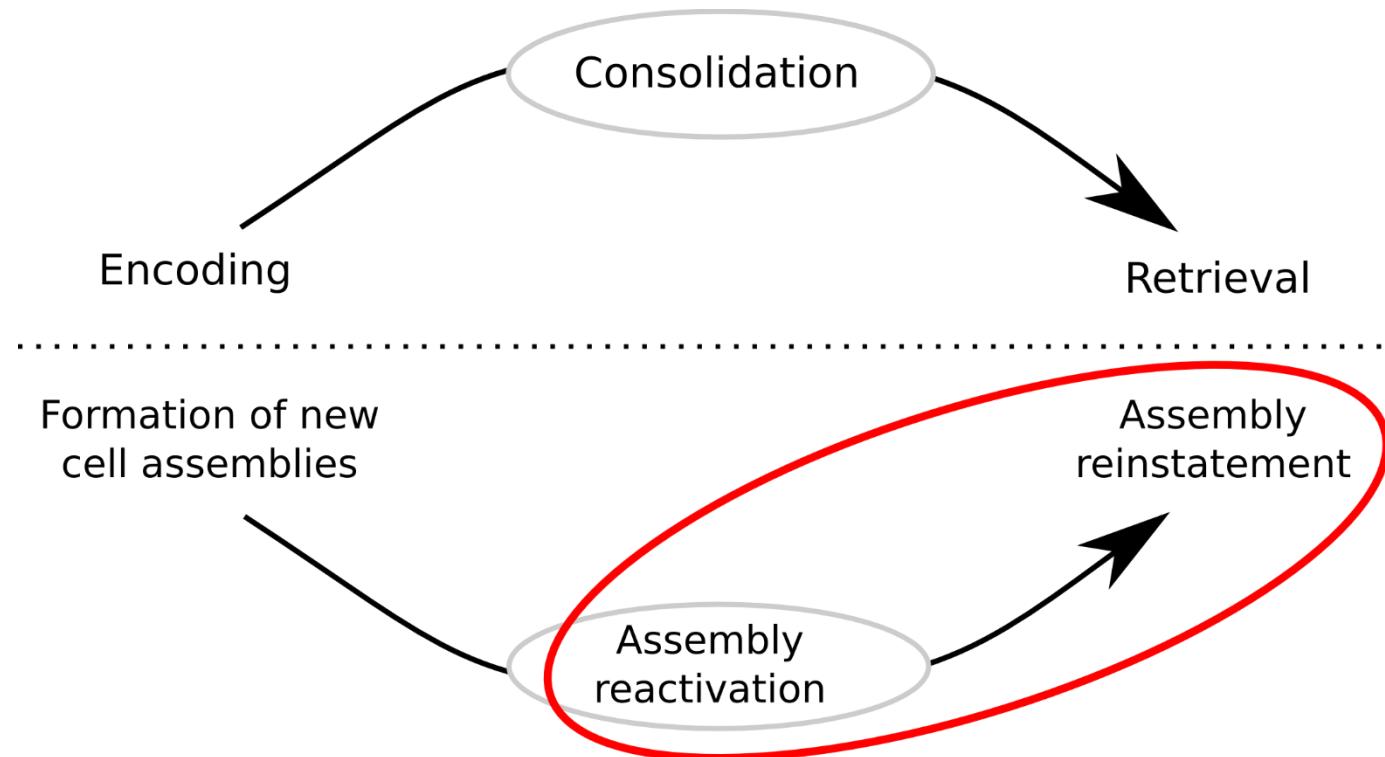
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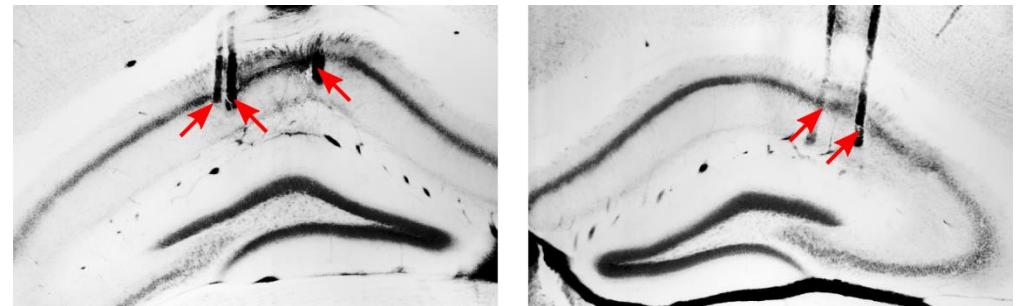
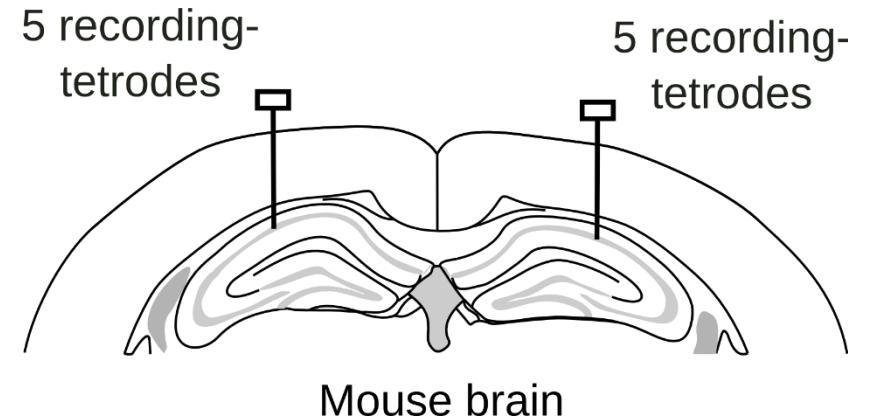
# Memory-consolidation: cell assembly / reactivation hypothesis



**2 challenges:** → Identification & tracking of “memory-representing” cell assemblies  
→ Selective disruption of reactivation

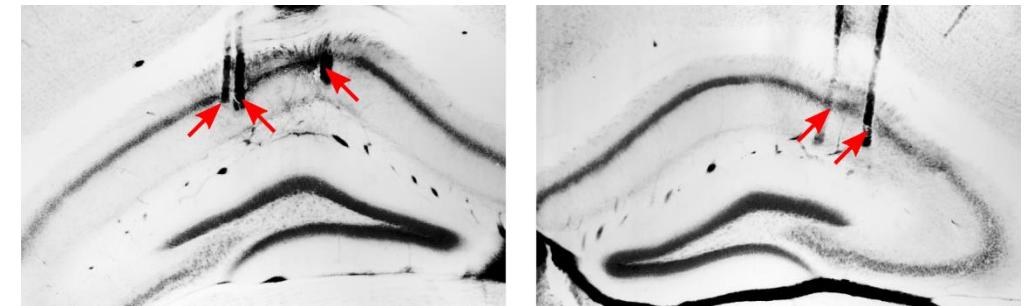
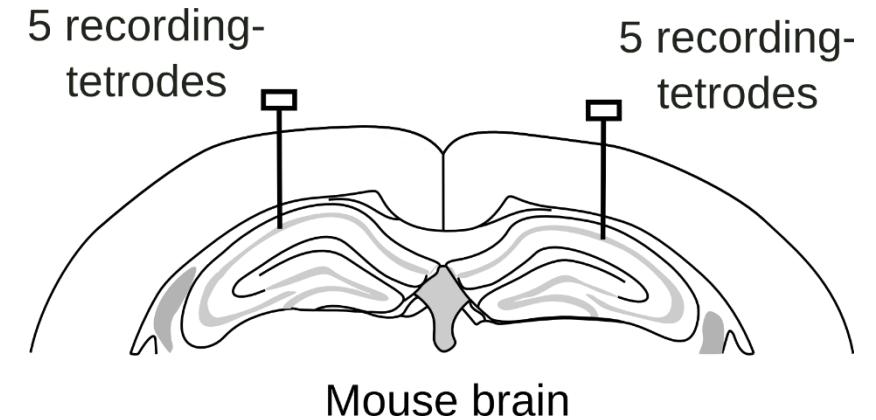
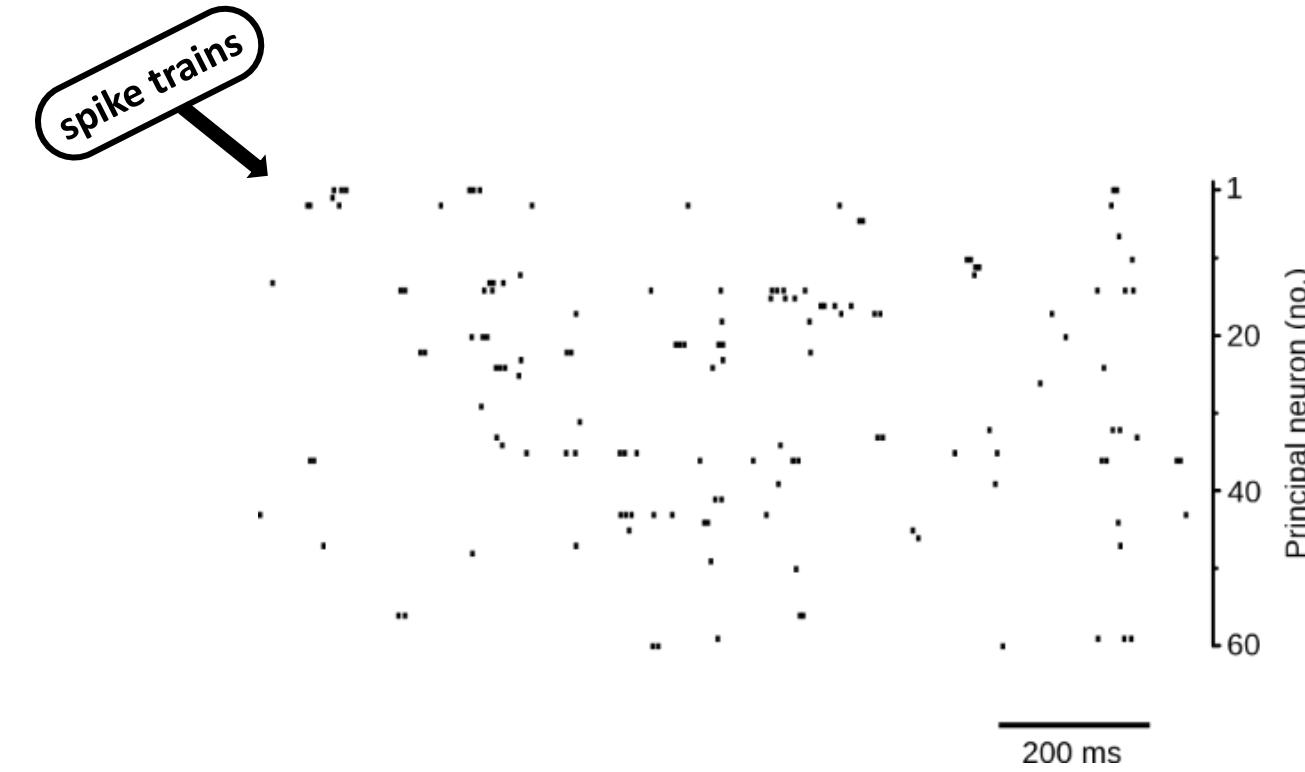
# Identification of “cell assemblies”

= challenge 1

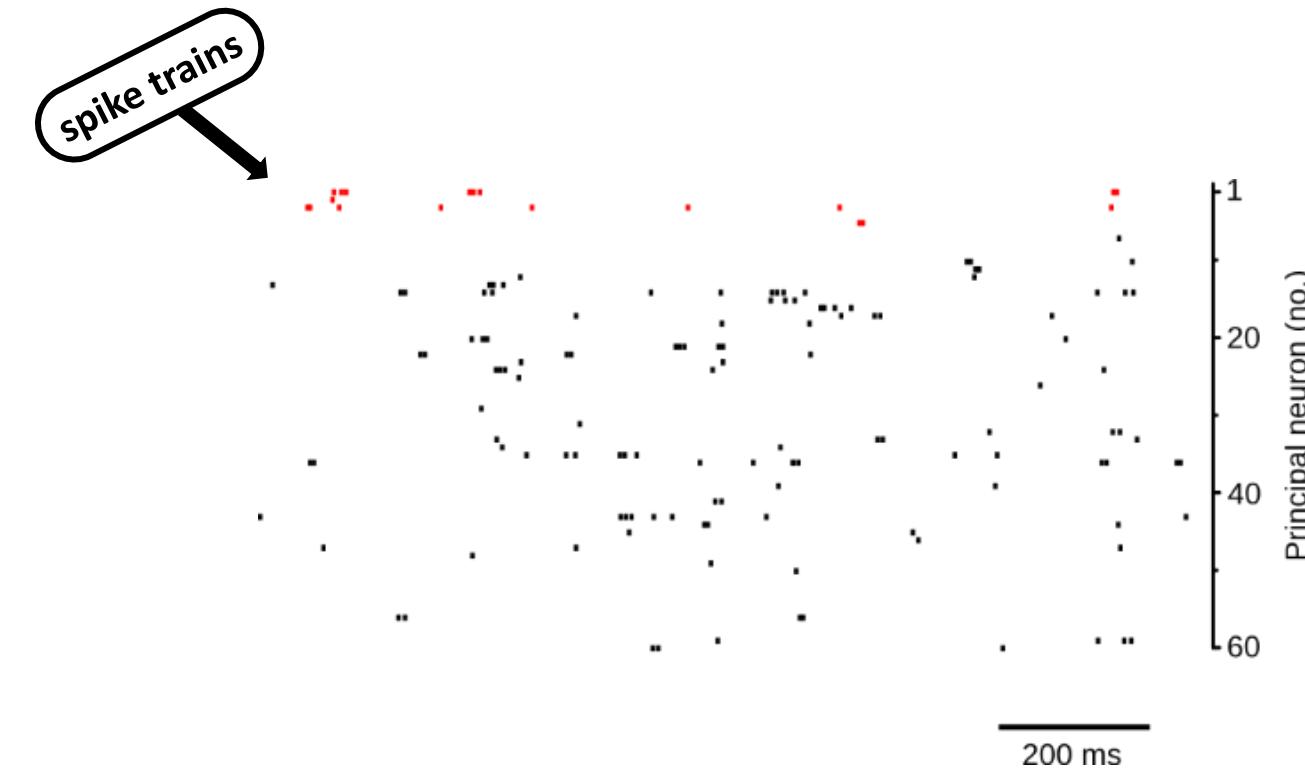


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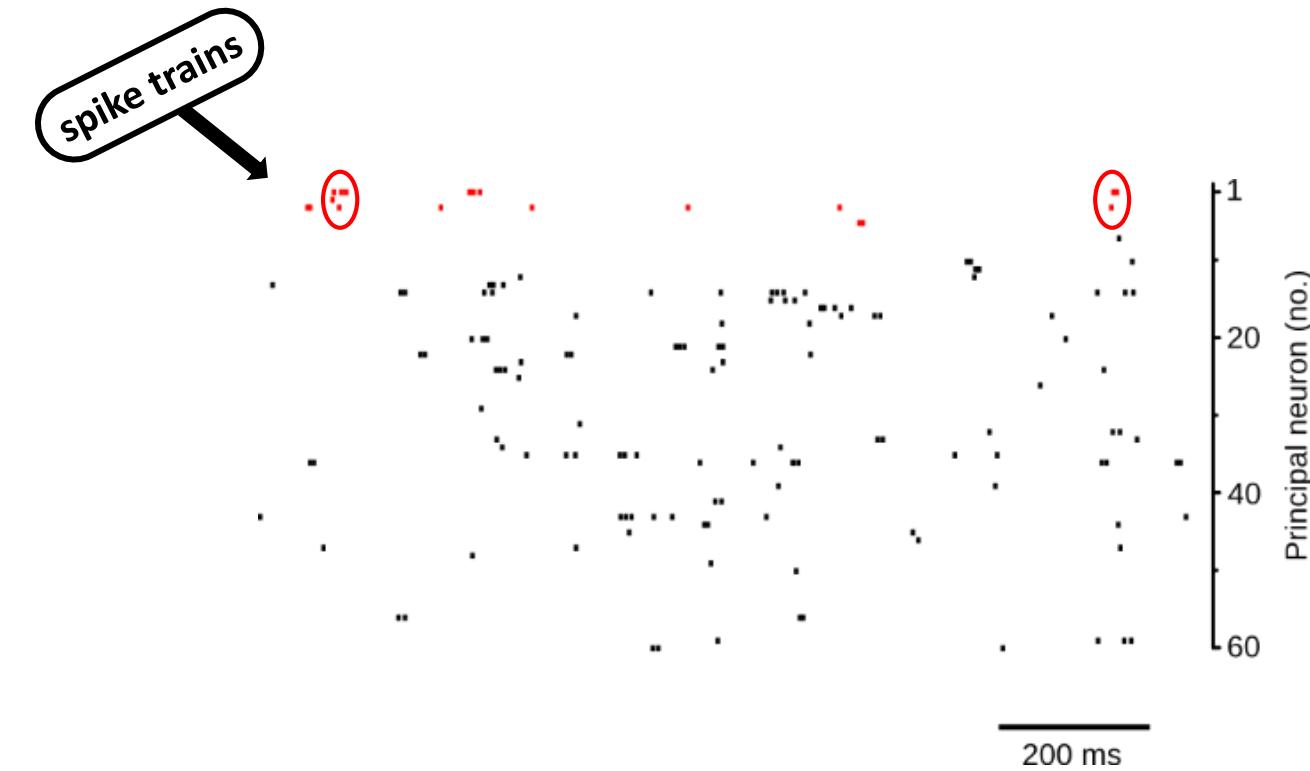
= challenge 1



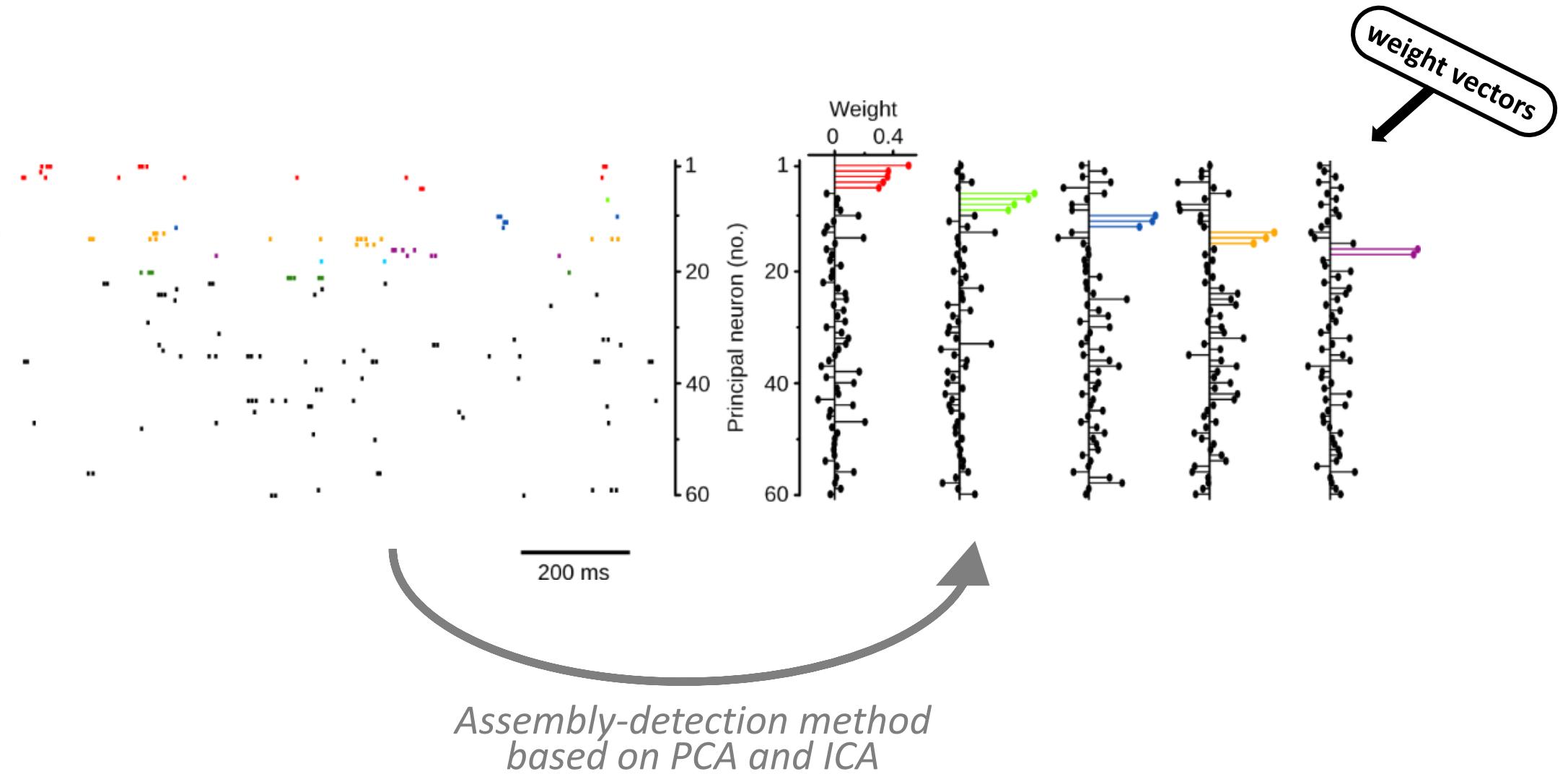
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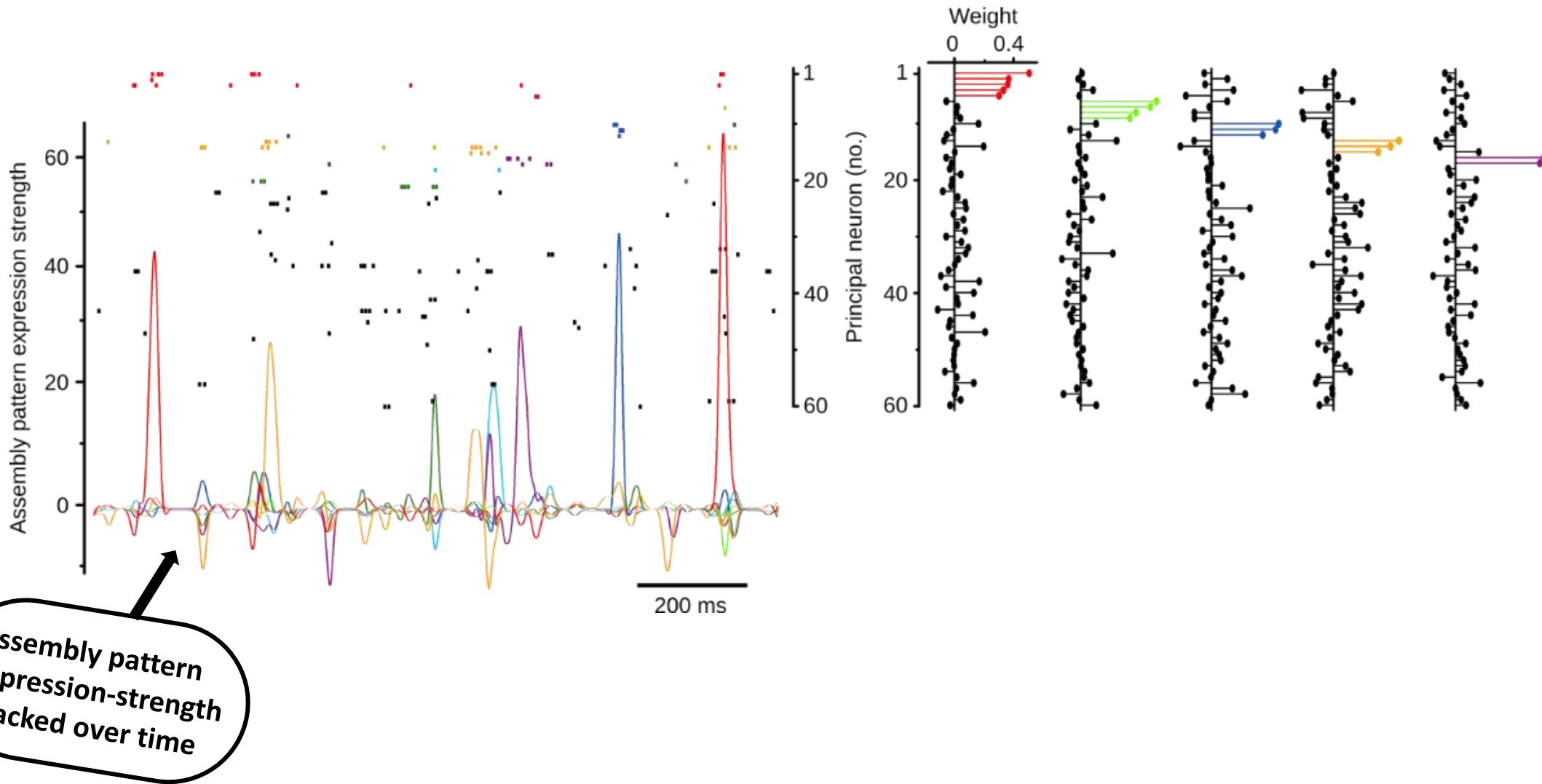
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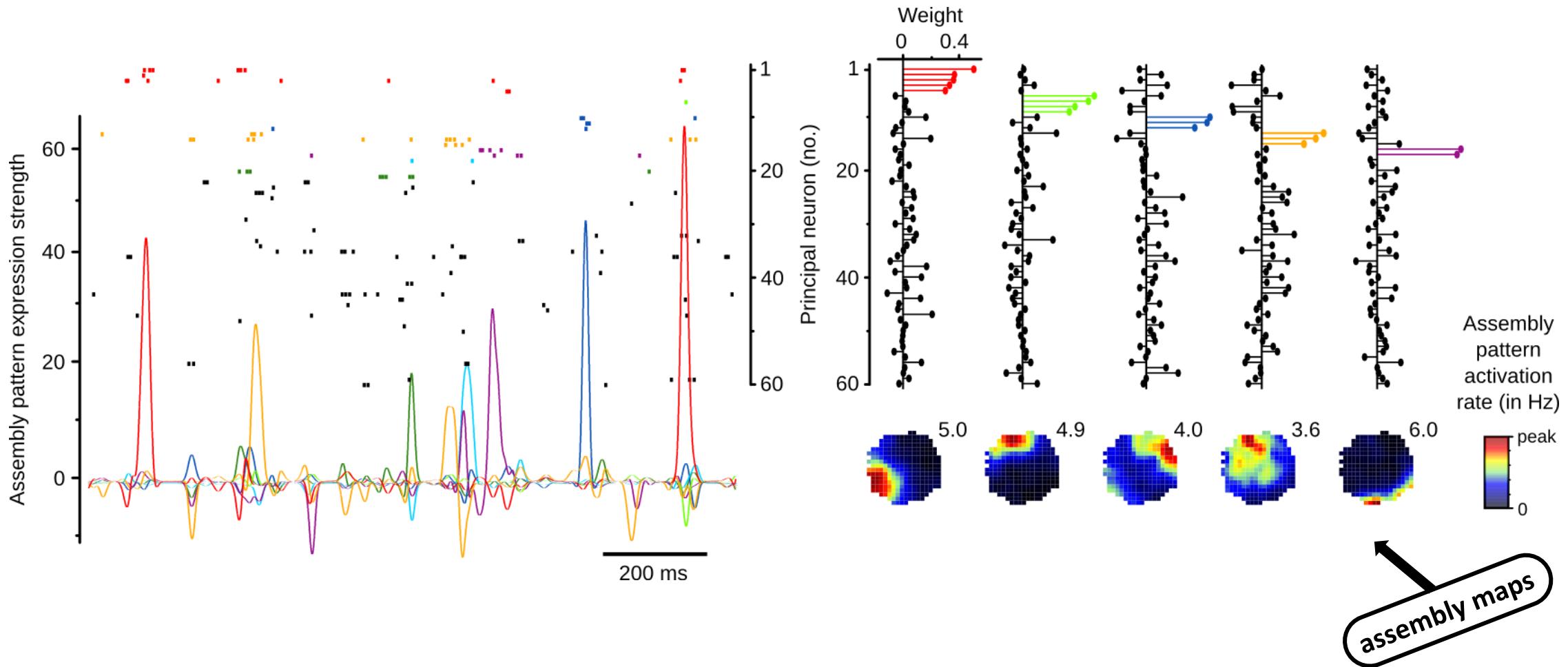
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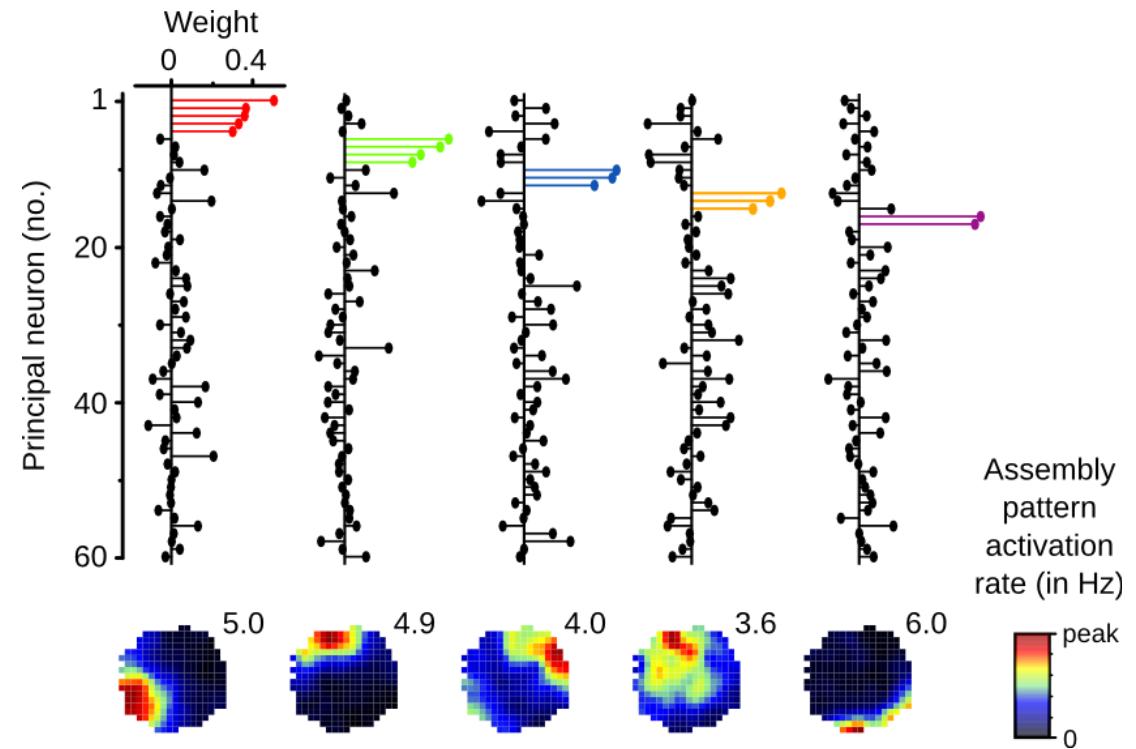
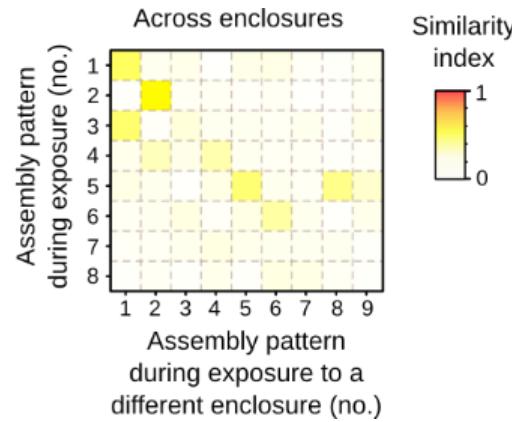
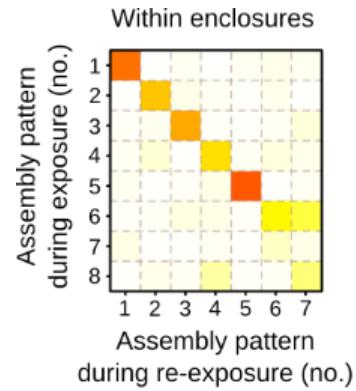


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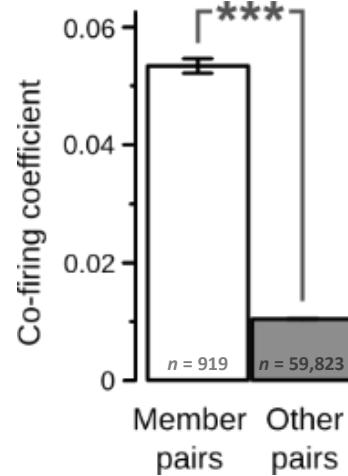
# Identification of “cell assemblies”

assembly patterns are environment-specific

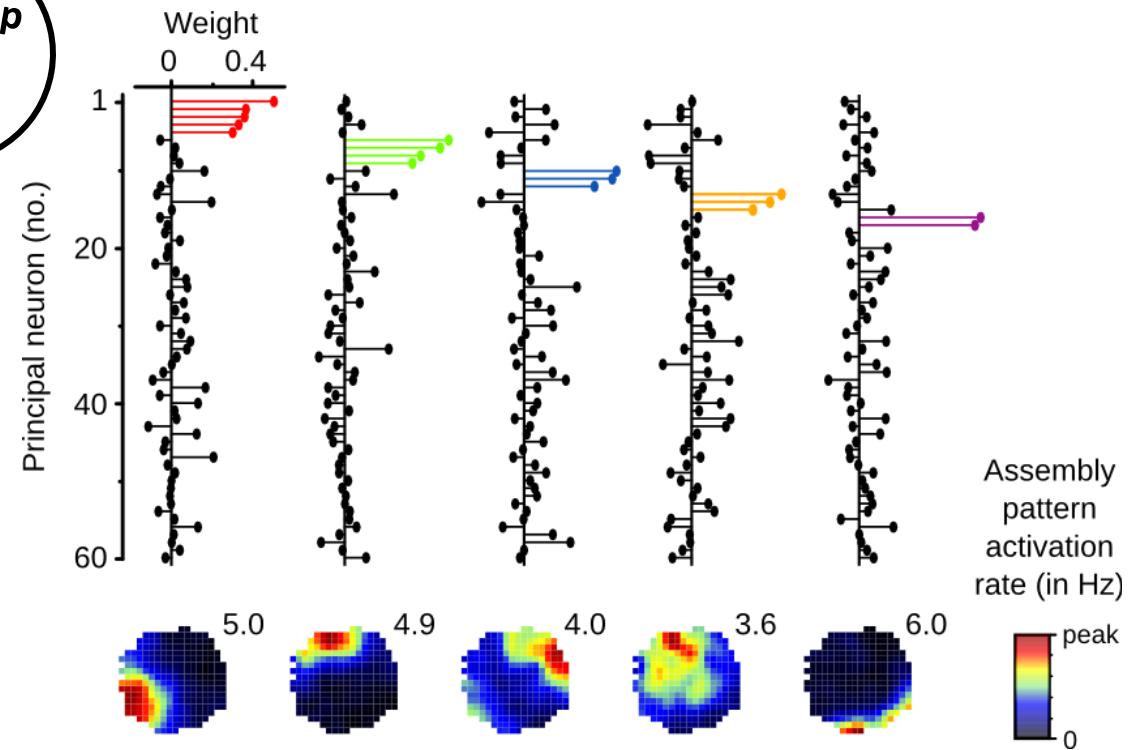
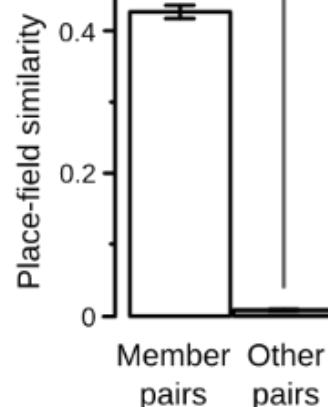


# Identification of “cell assemblies”

assembly patterns group together co-active neurons

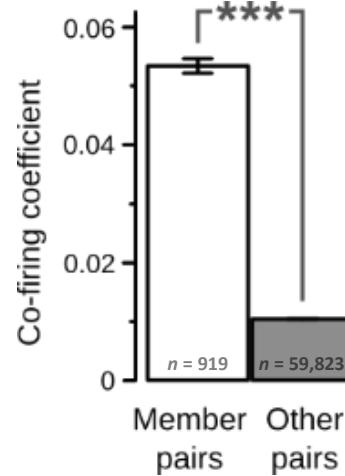


assembly patterns group together neurons with overlapping place fields

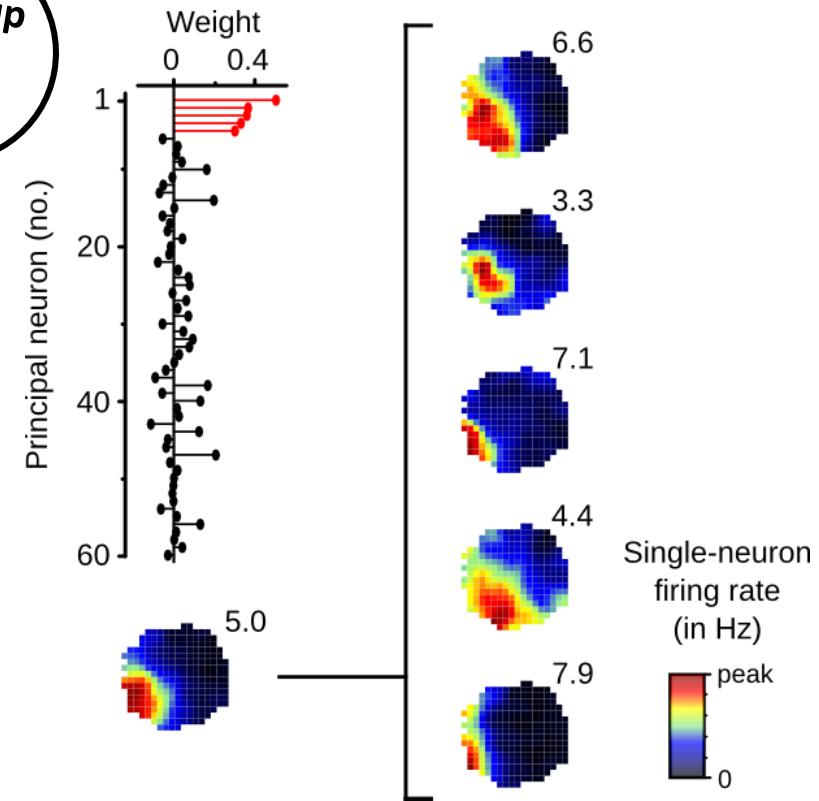
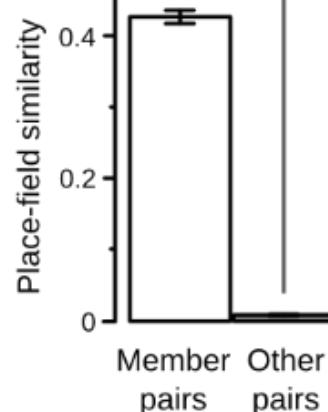


# Identification of “cell assemblies”

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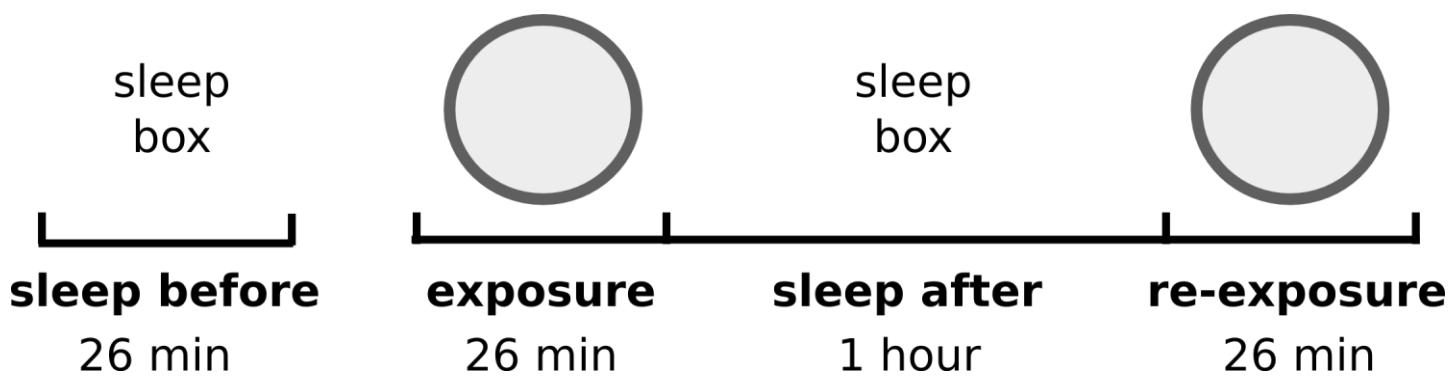
# Identification of “cell assemblies” - summary

Based on short-time scale interactions (25 ms) in tetrode recordings of hippocampal principal neurons, an unsupervised statistical framework based on PCA and ICA ***detects*** and ***tracks*** cell assembly patterns that:

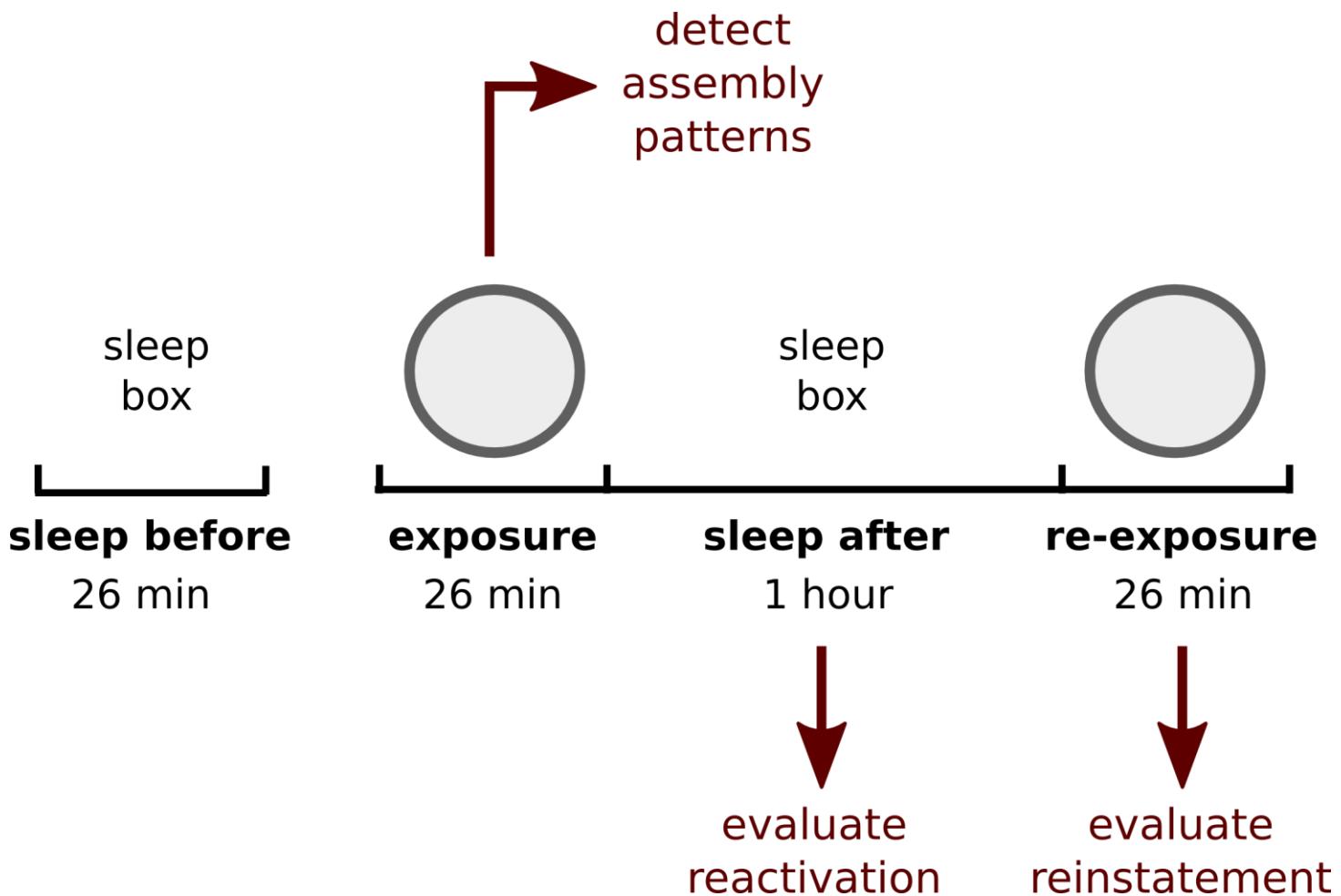
- are spatially selective;
- are environment-specific;
- bind together co-active neurons; and
- bind together neurons with overlapping spatial tuning.

→ **challenge 1:** Identification & tracking of “memory-representing” cell assemblies

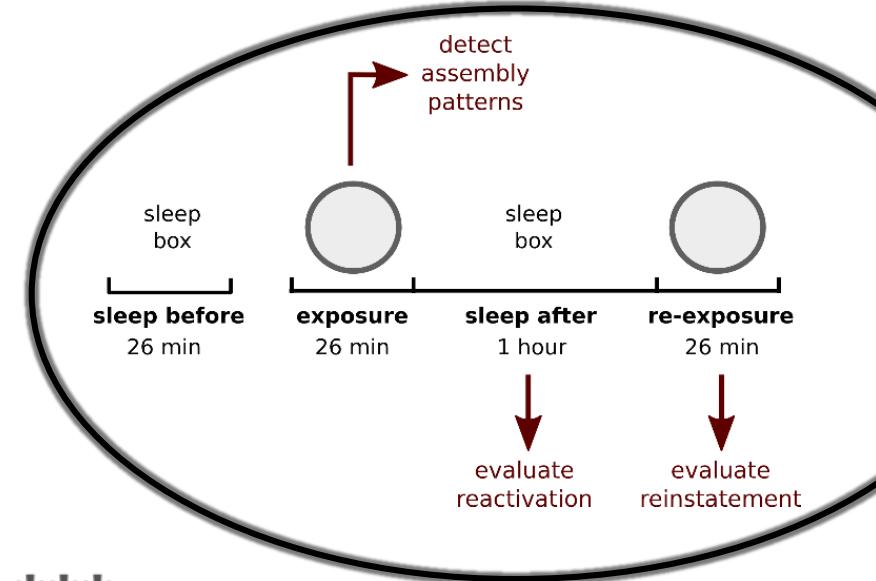
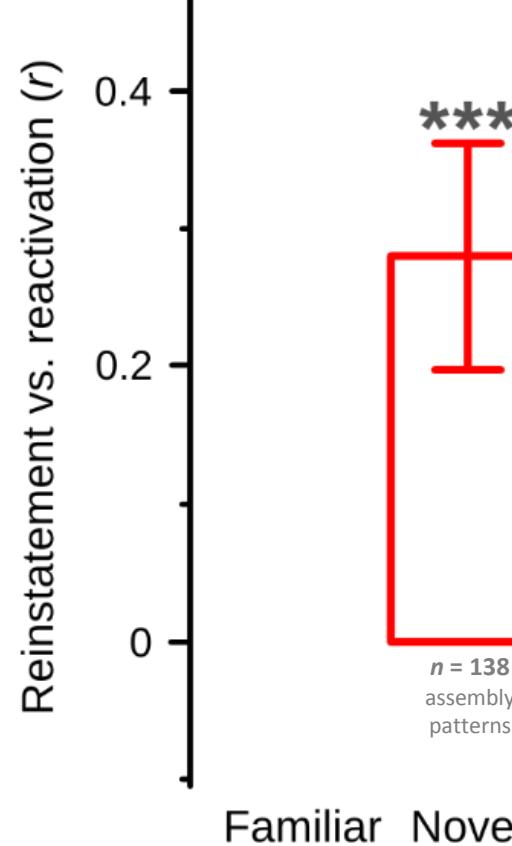
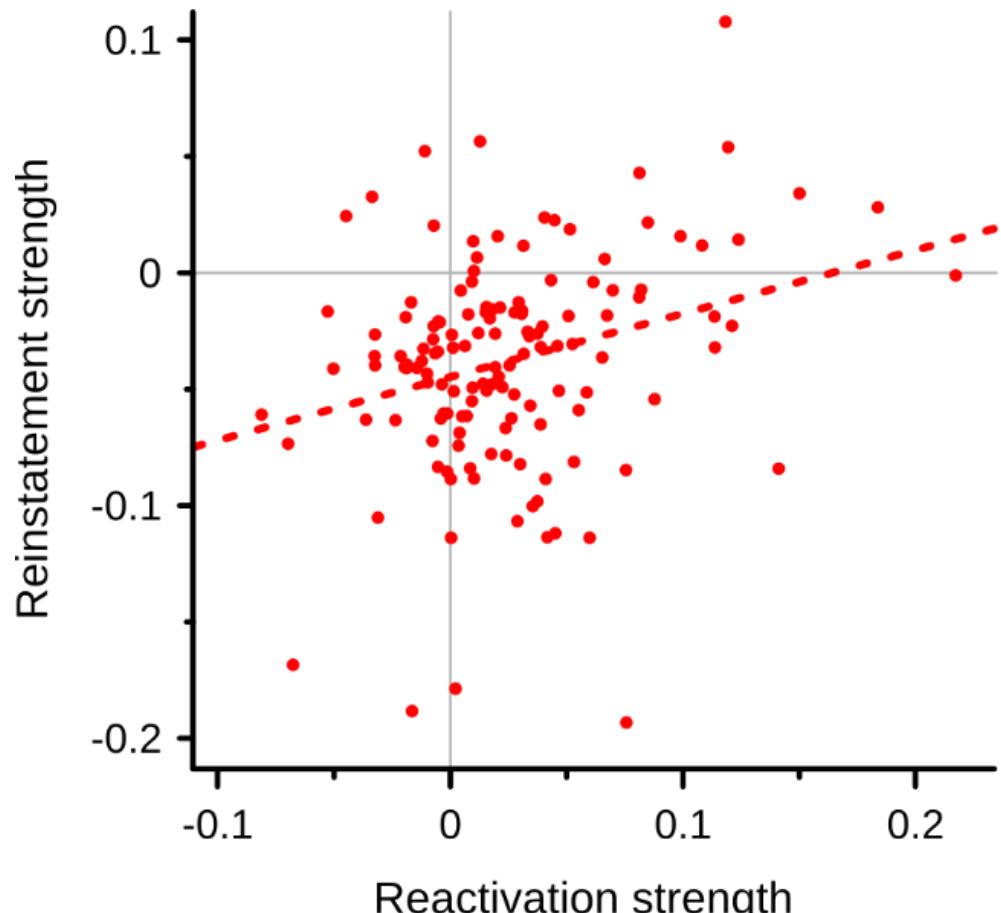
# Experimental protocol



# Experimental protocol - (correlation)

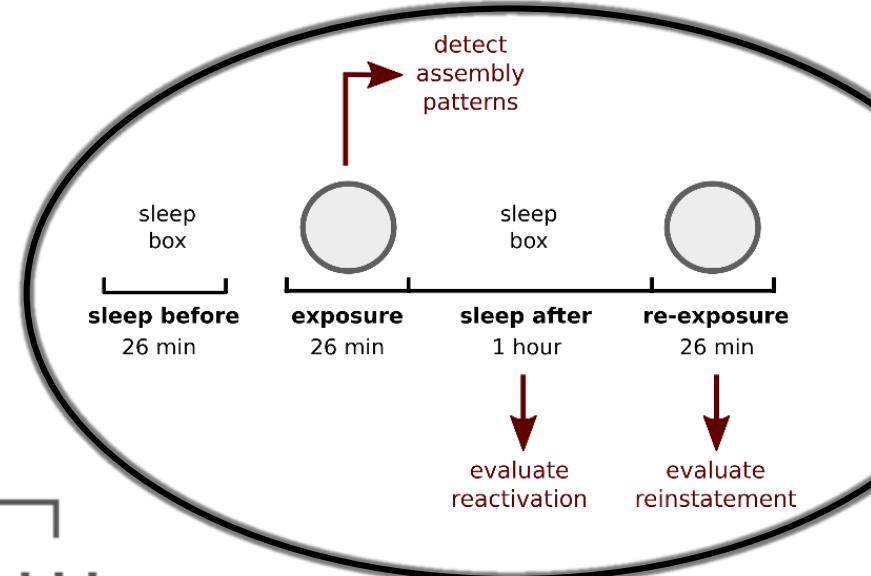
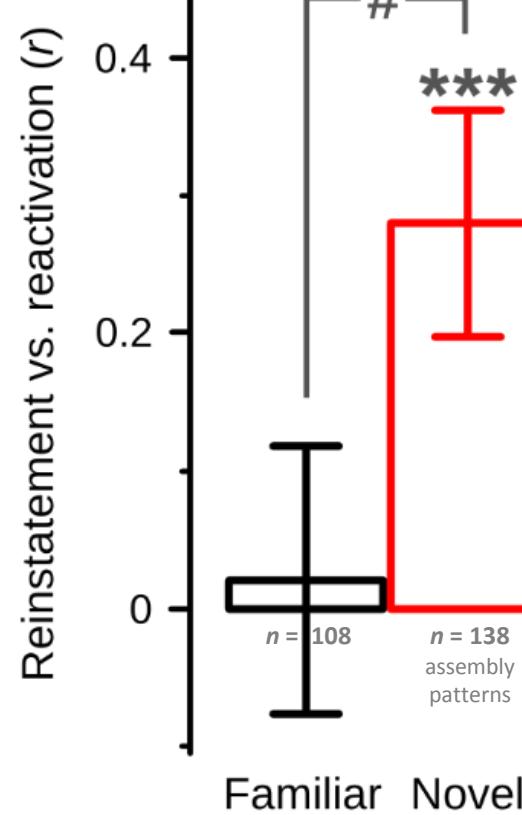
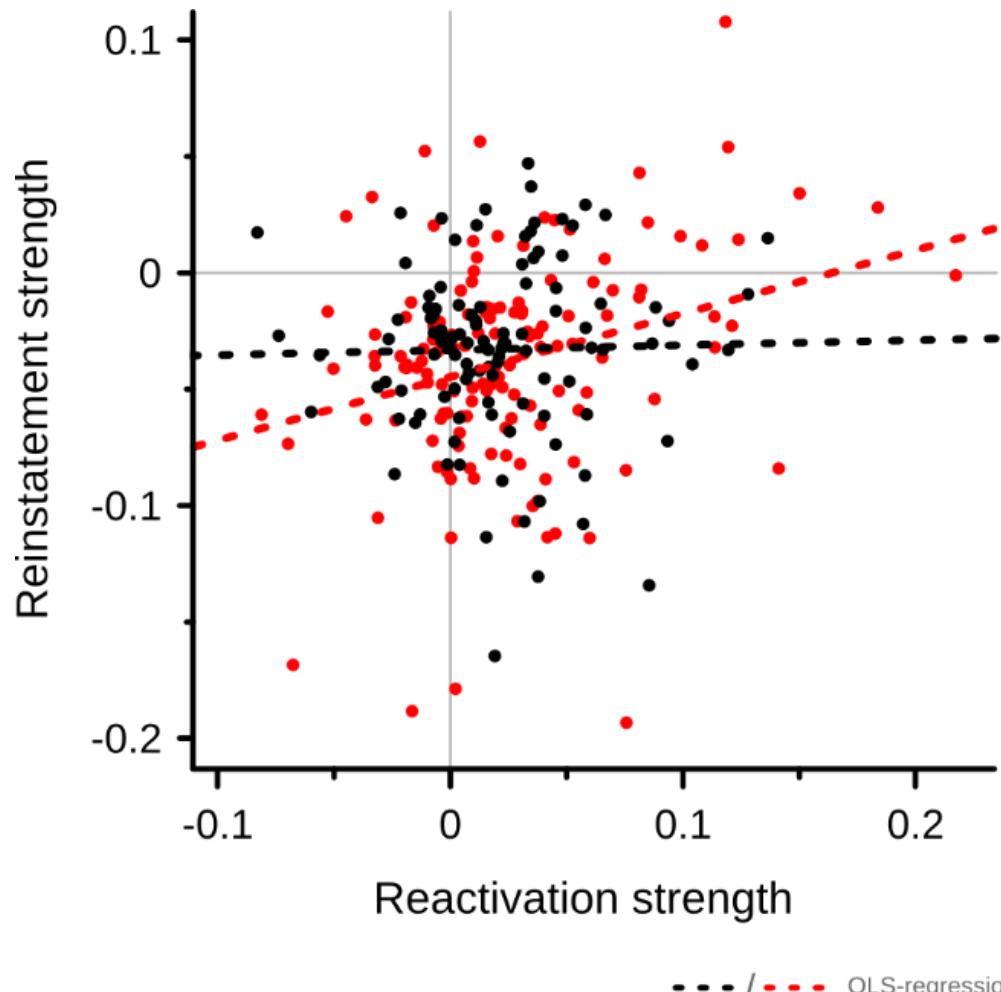


# An assembly pattern's reactivation predicts its subsequent reinstatement



(based on 43 recording-blocks from 8 mice)

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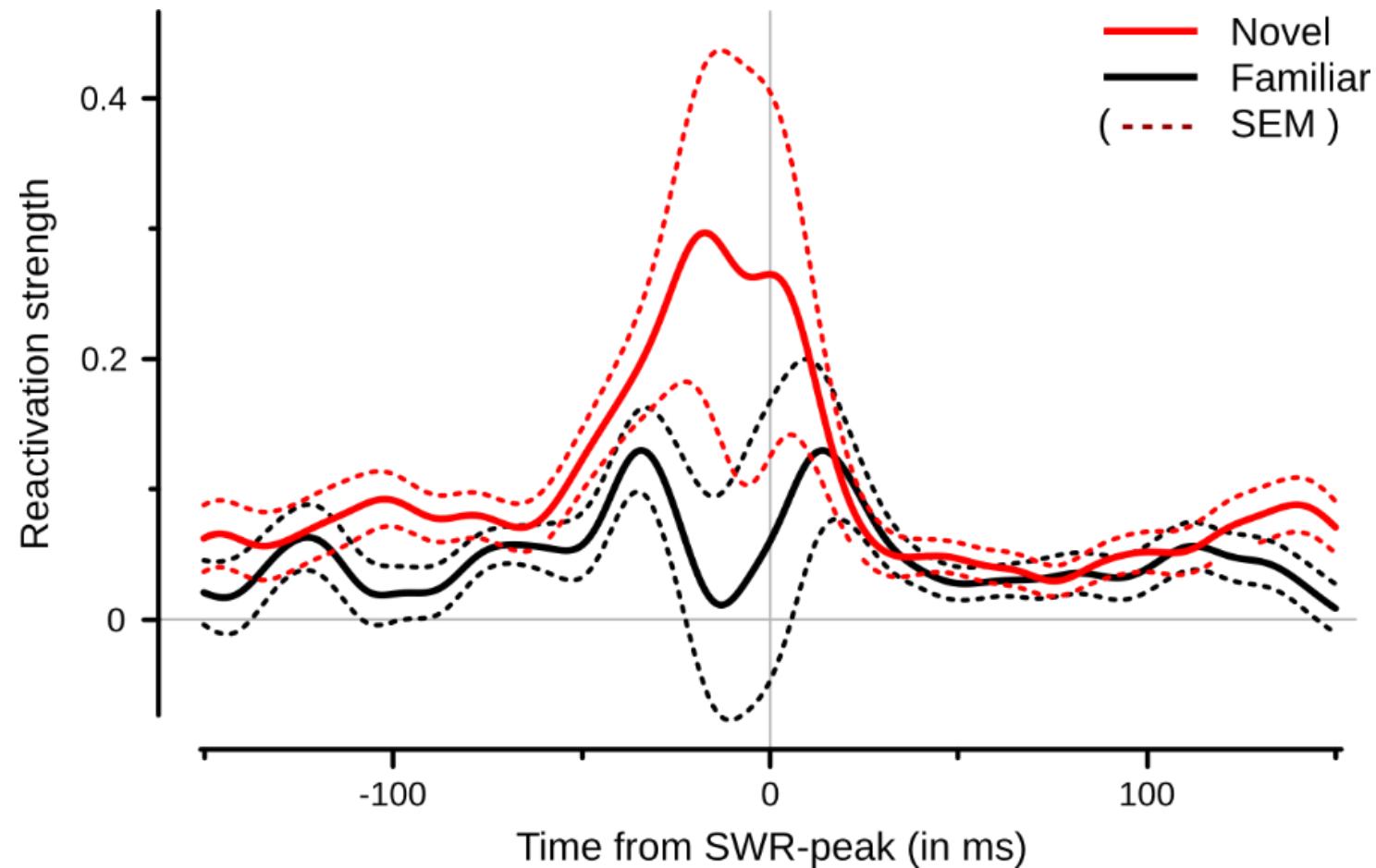
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Selective disruption of reactivation?

= challenge 2

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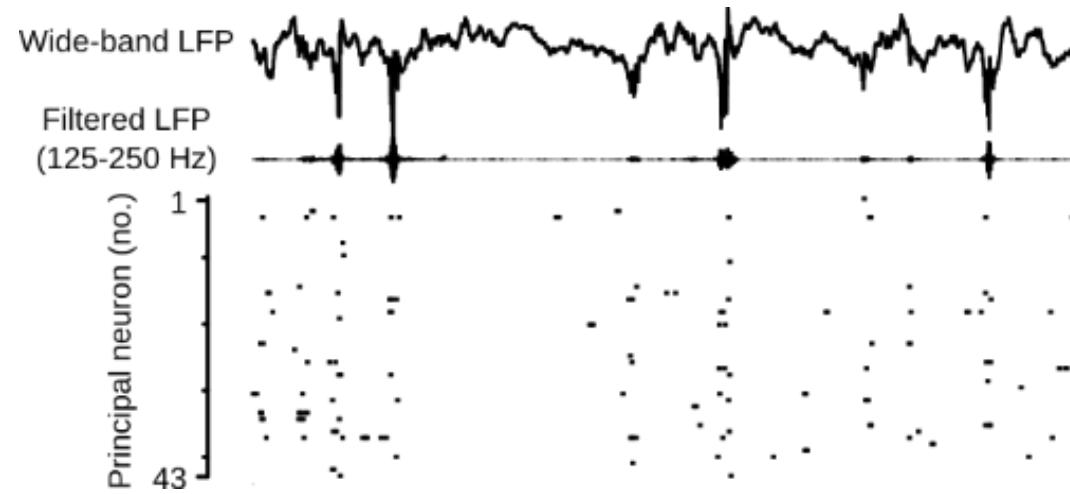


Novel:  $n = 139$  assembly-patterns

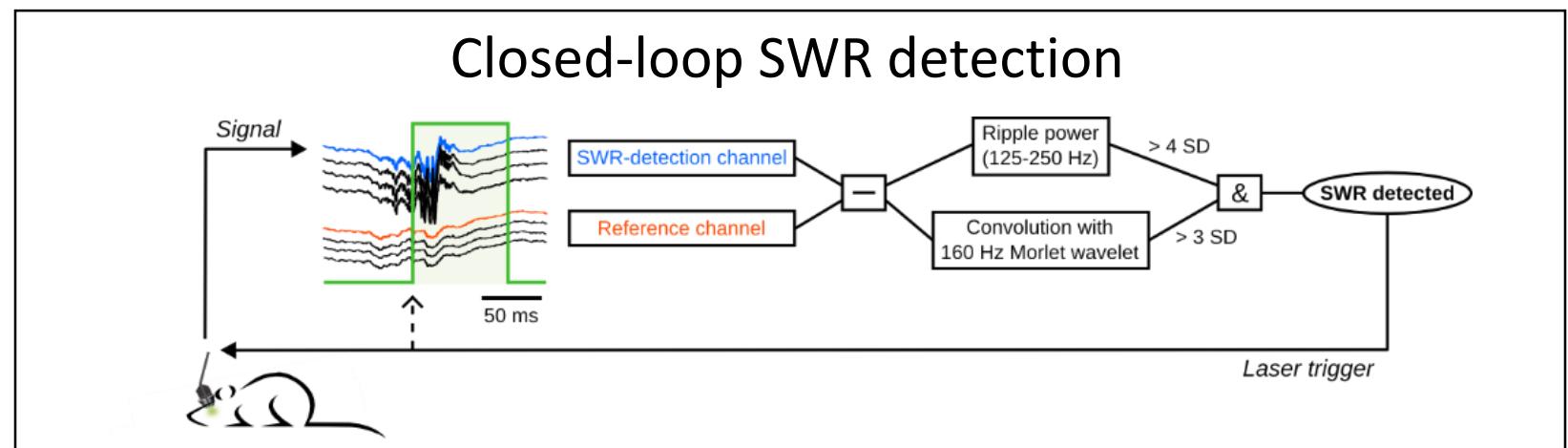
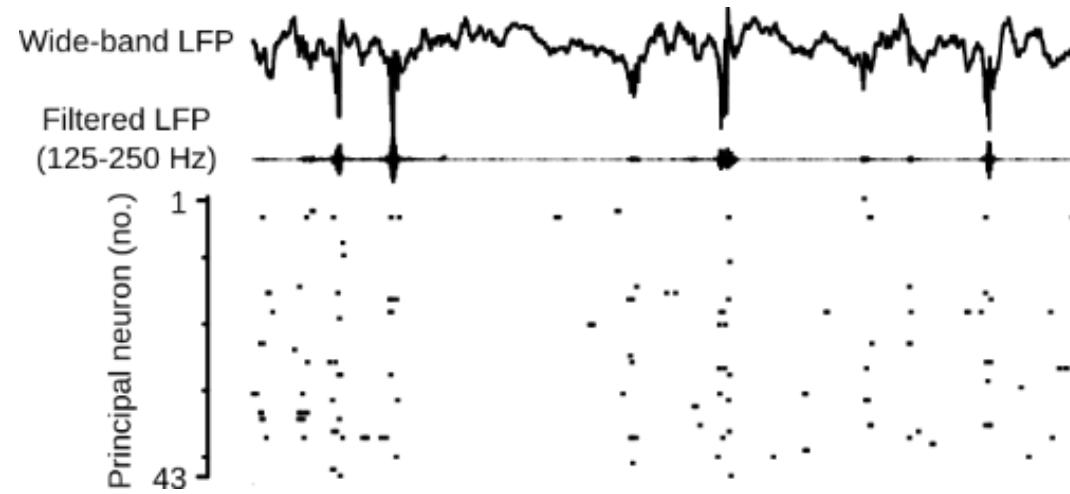
Familiar:  $n = 108$  assembly-patterns

(based on 43 recording-blocks from 8 mice)

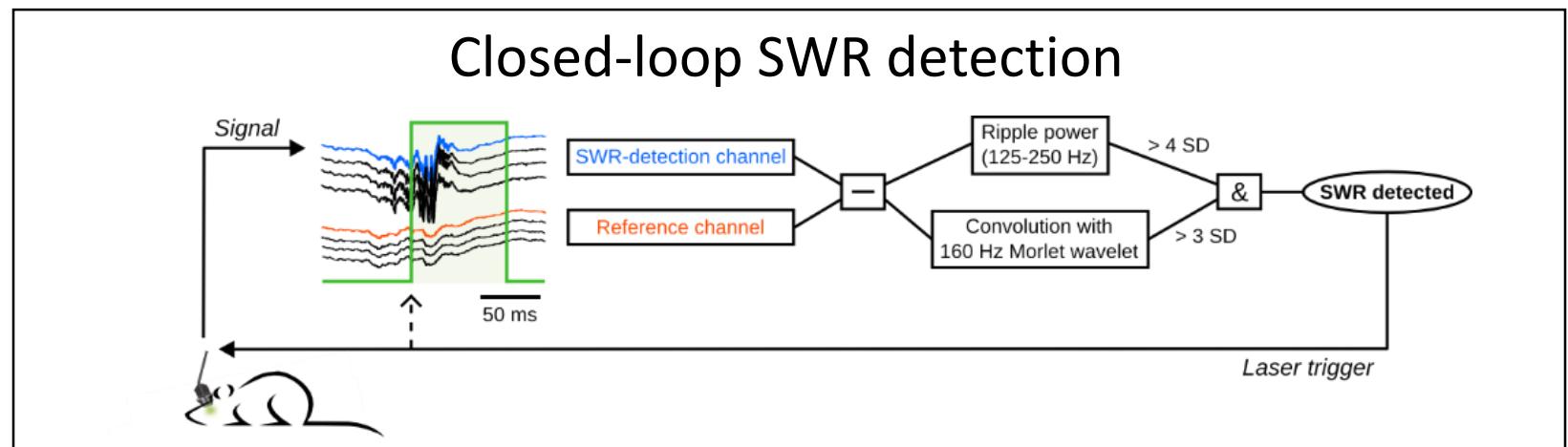
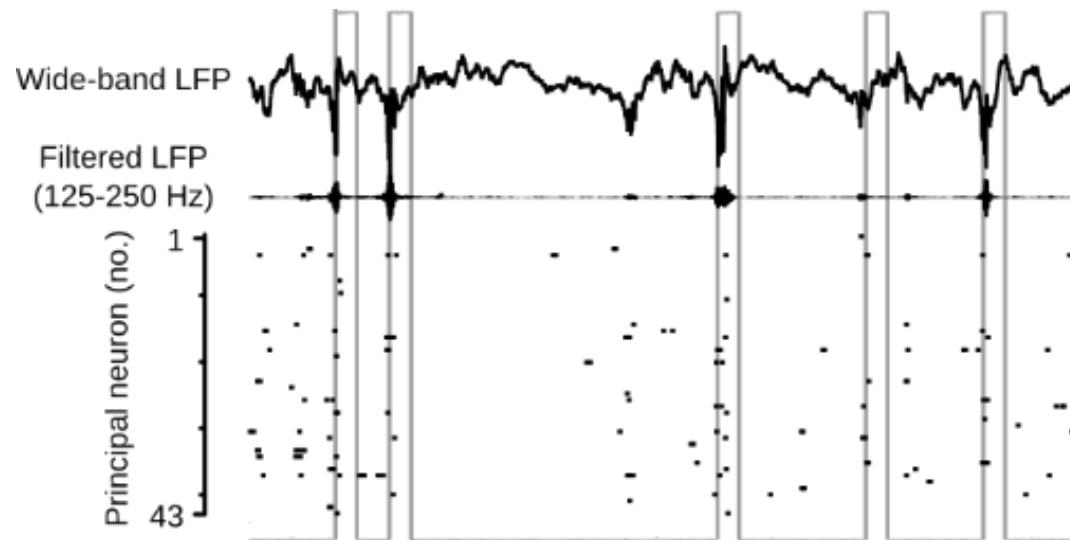
# Selective disruption of reactivation: *optogenetic SWR silencing*



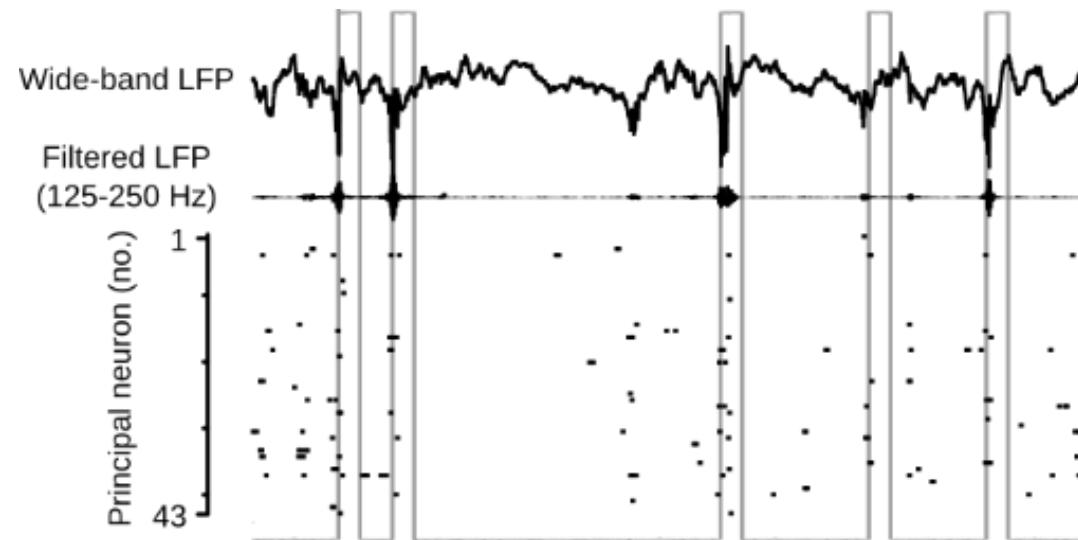
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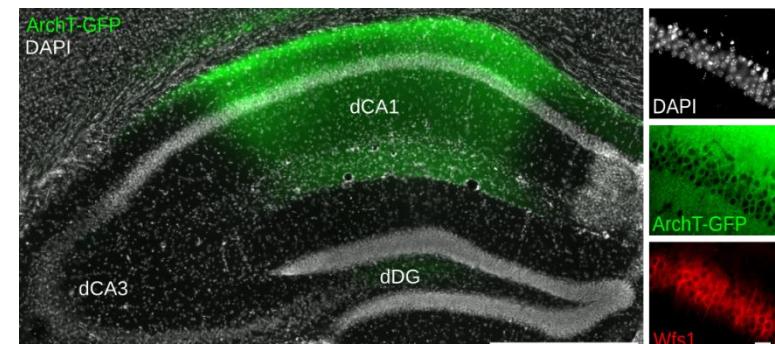
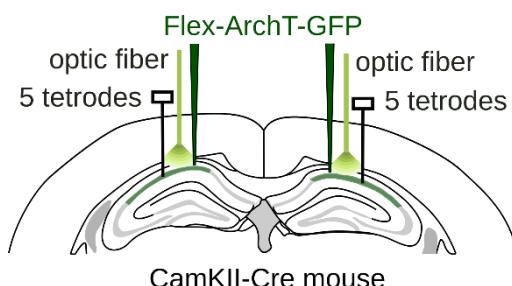
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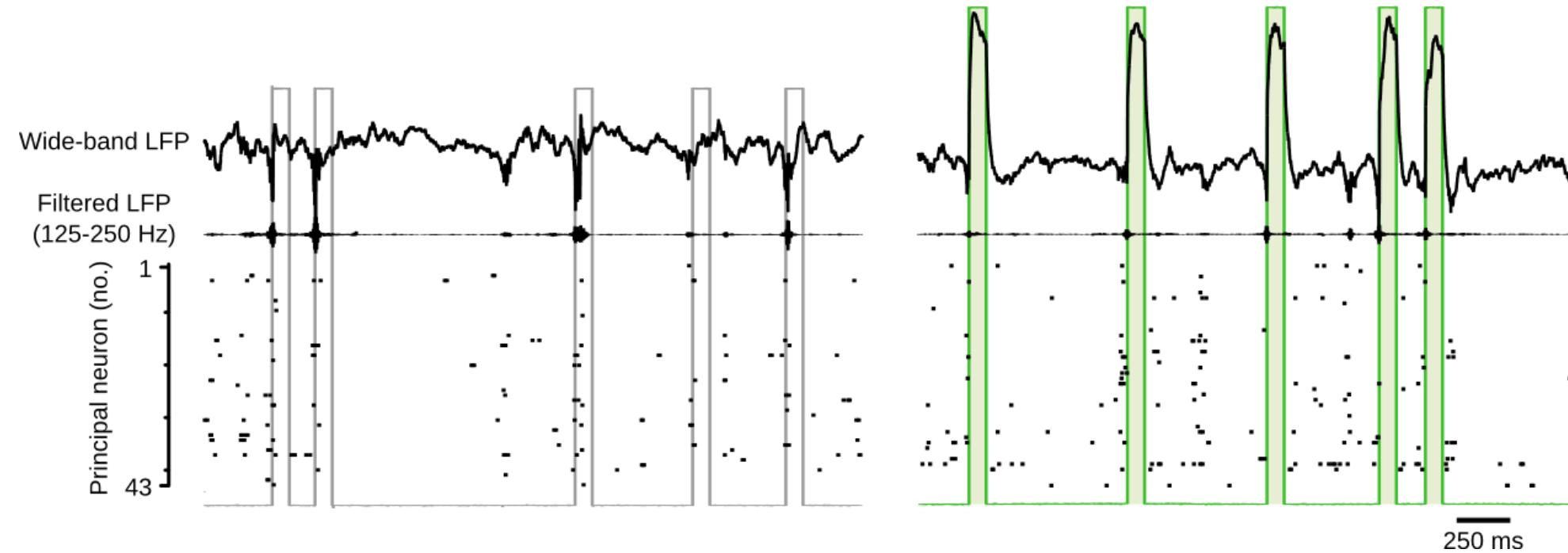
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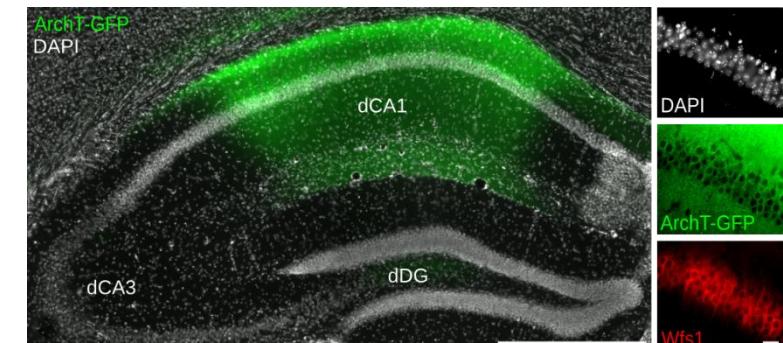
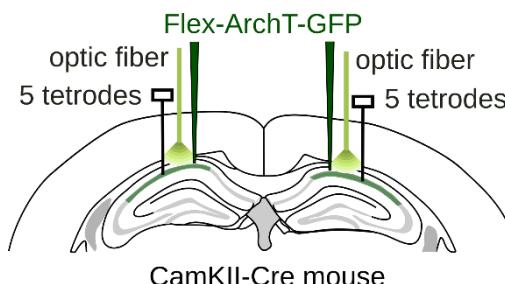
## Optogenetic silencing of CamKII-positive cells using ArchT



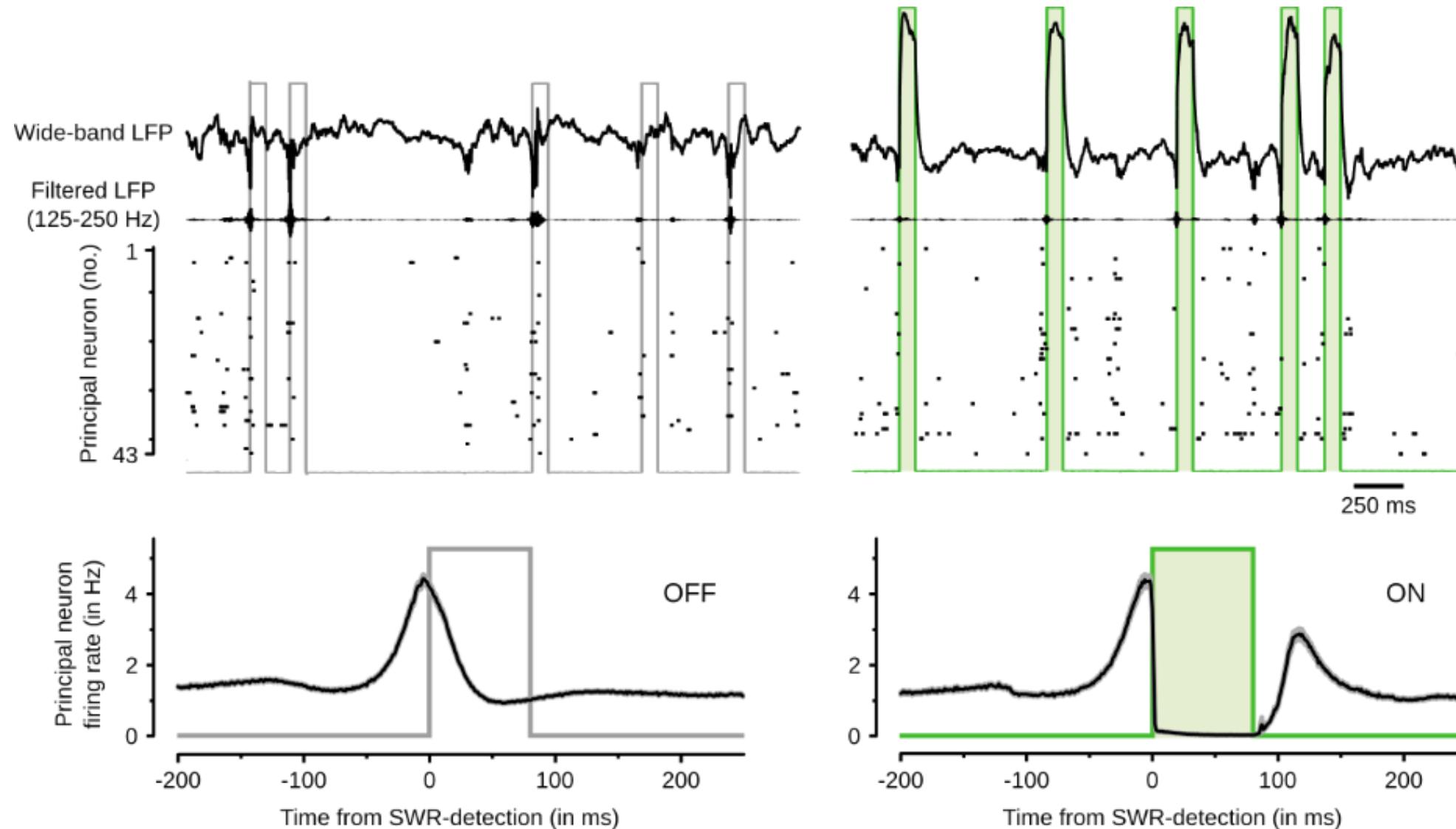
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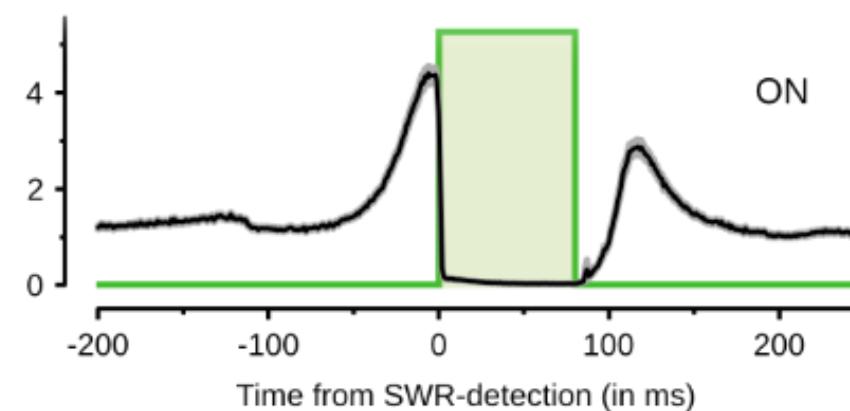
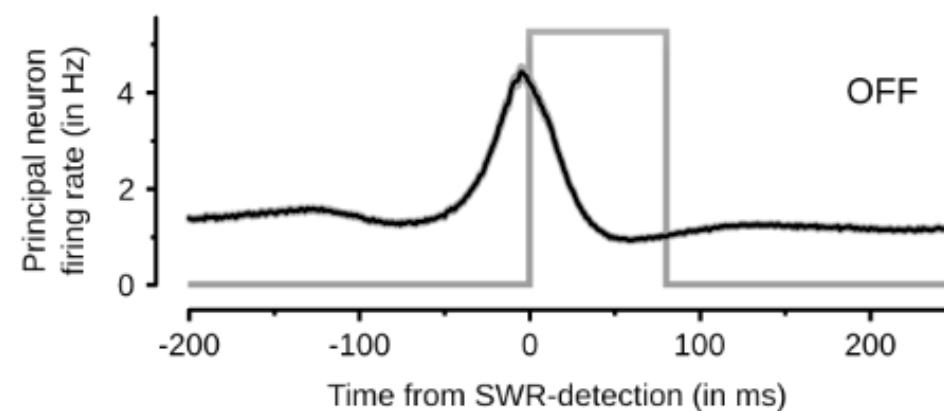
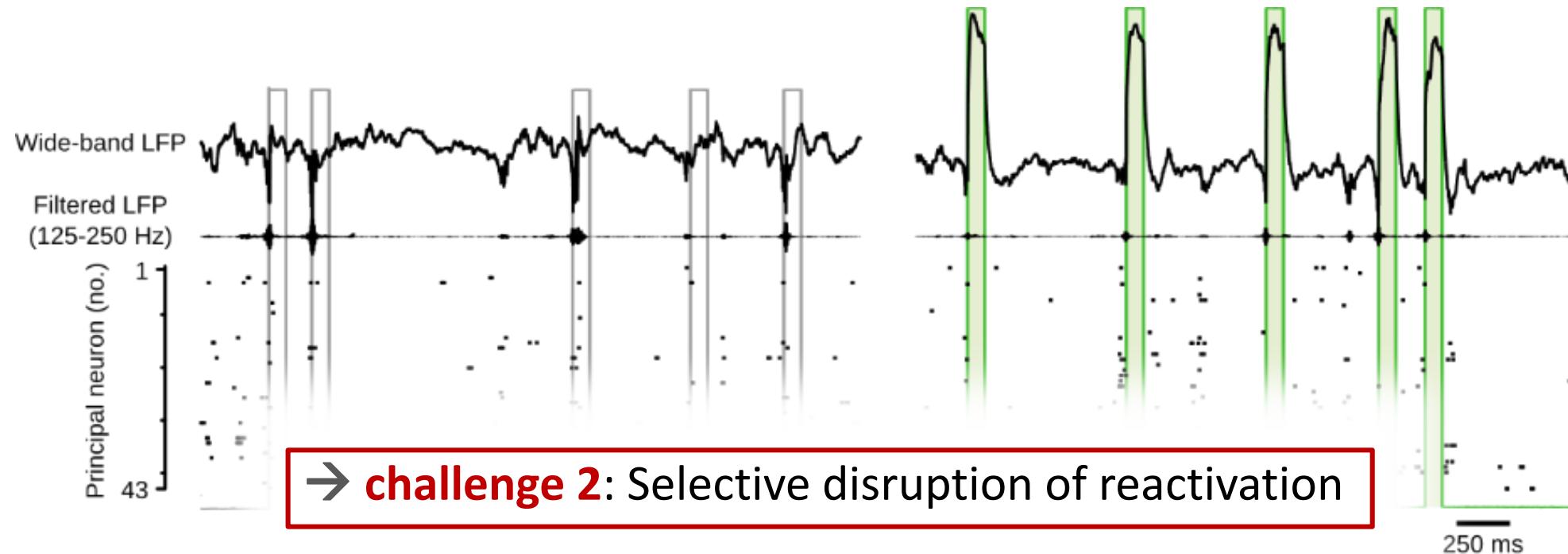


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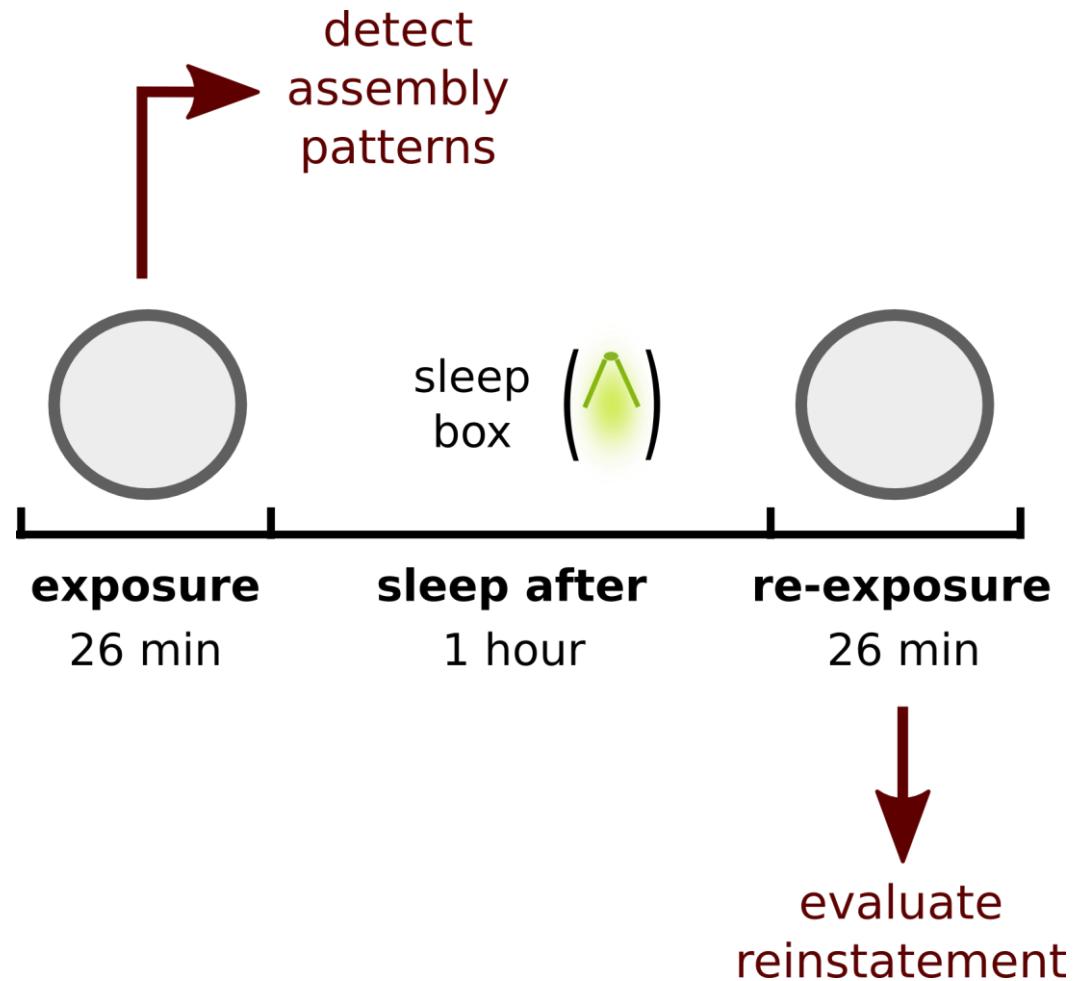
OFF:  $n = 1,988$  neurons (from 43 sessions)  
ON:  $n = 1,527$  neurons (from 37 sessions)

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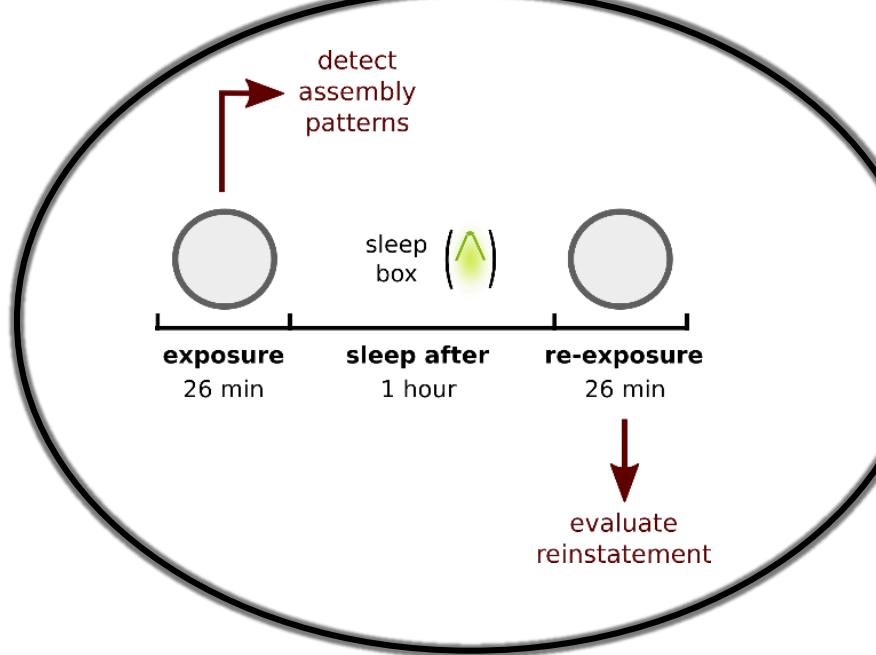
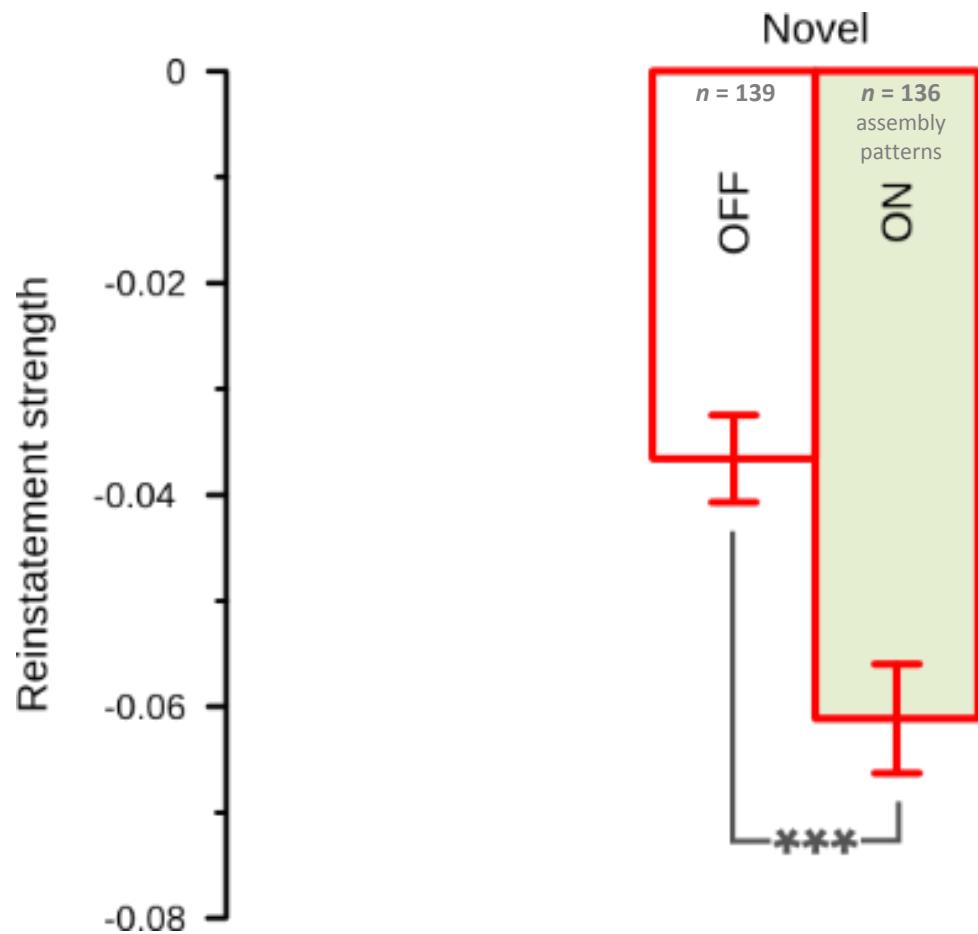


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# Experimental protocol - (*causation*)

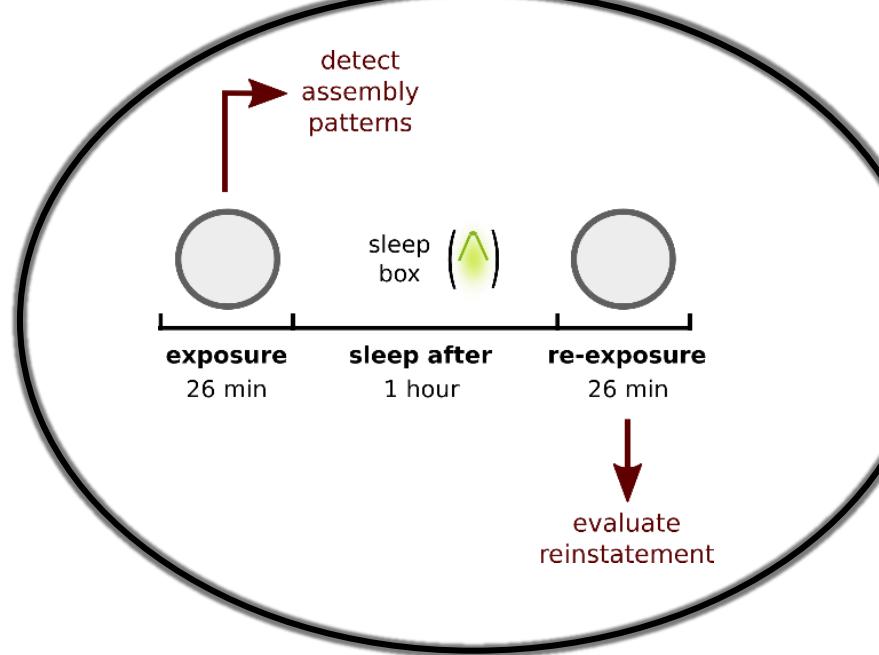
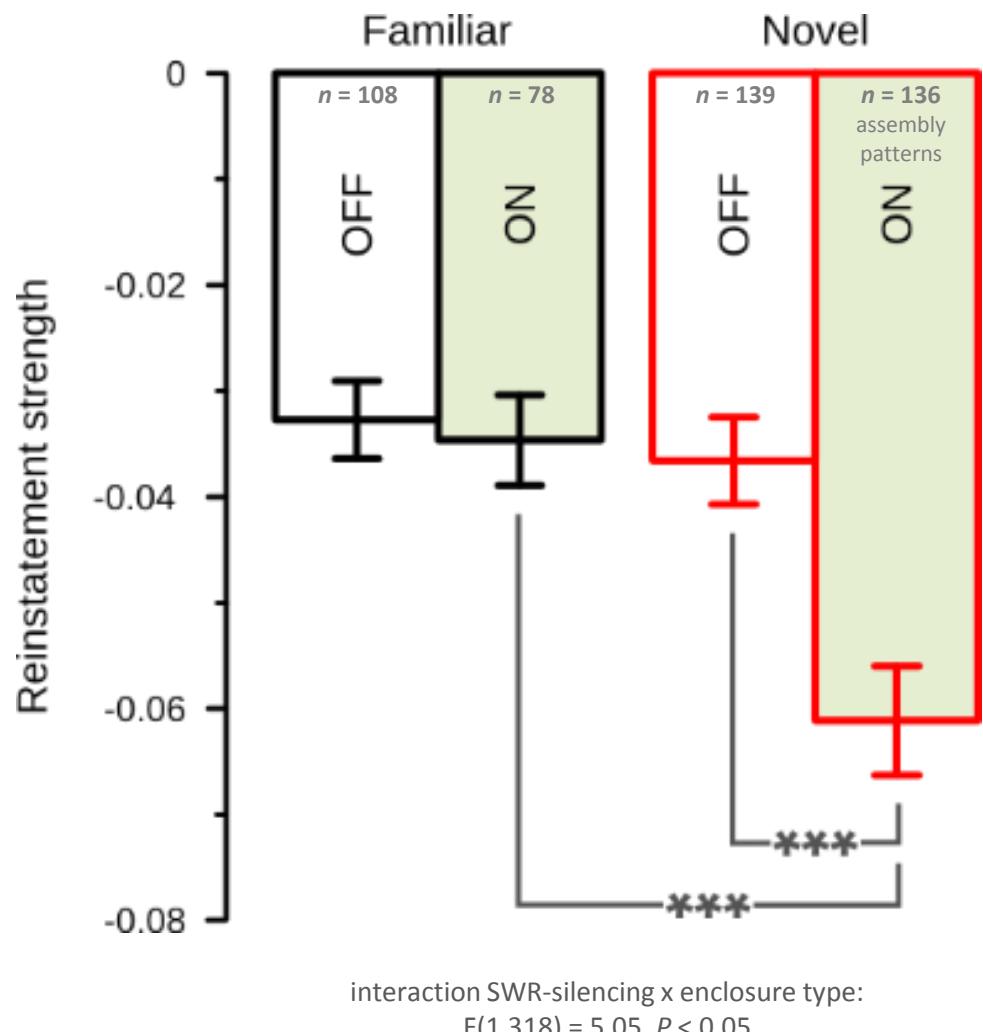


# SWR-silencing impairs assembly pattern reinstatement



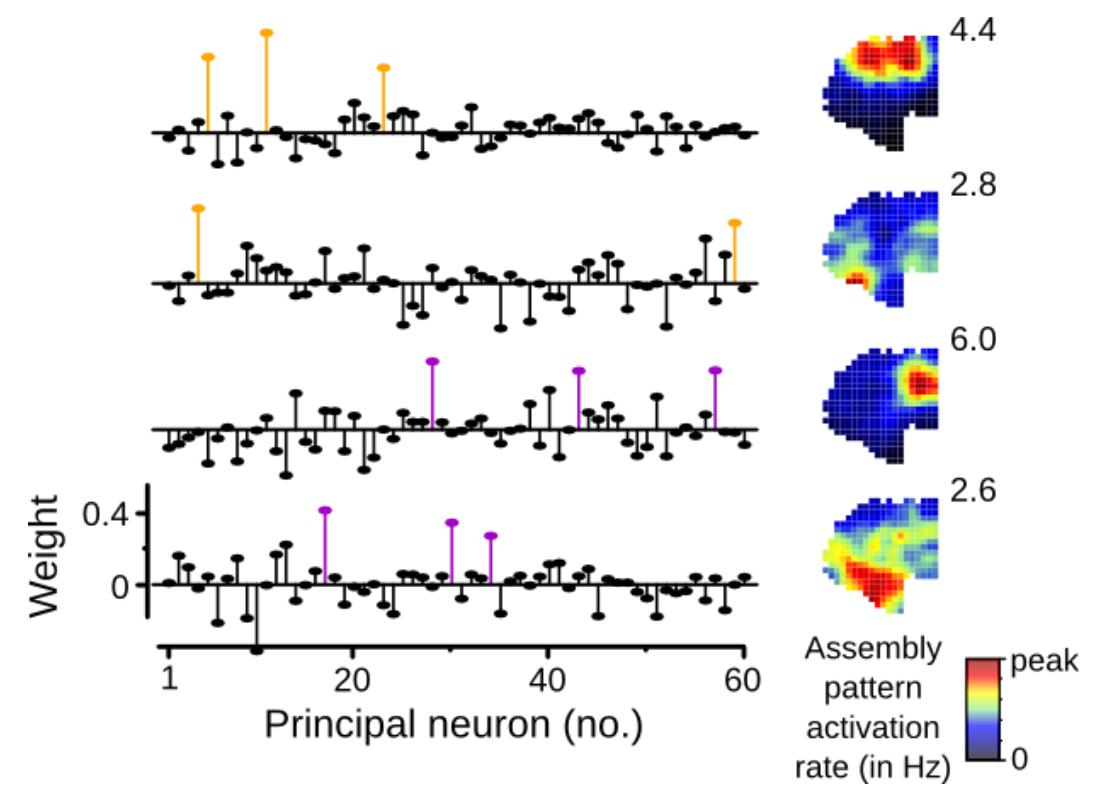
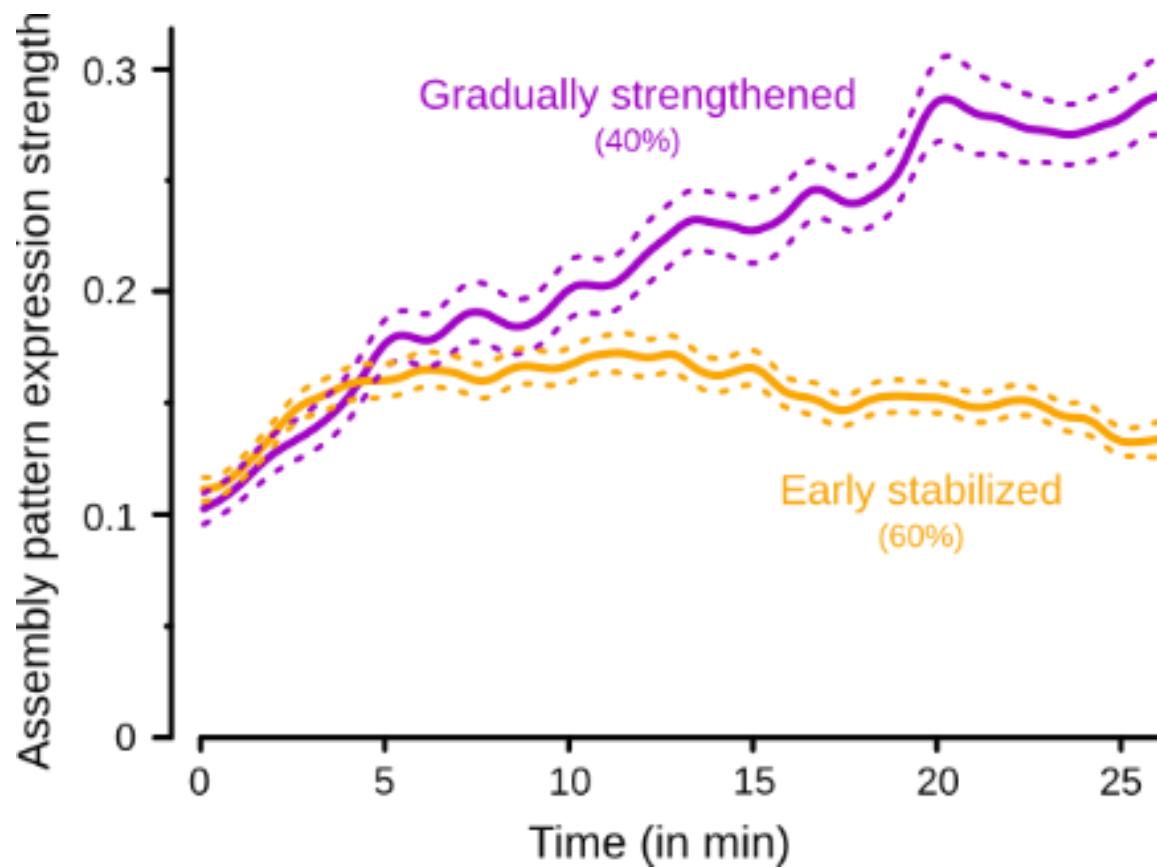
(based on 50 recording-blocks from 8 mice)

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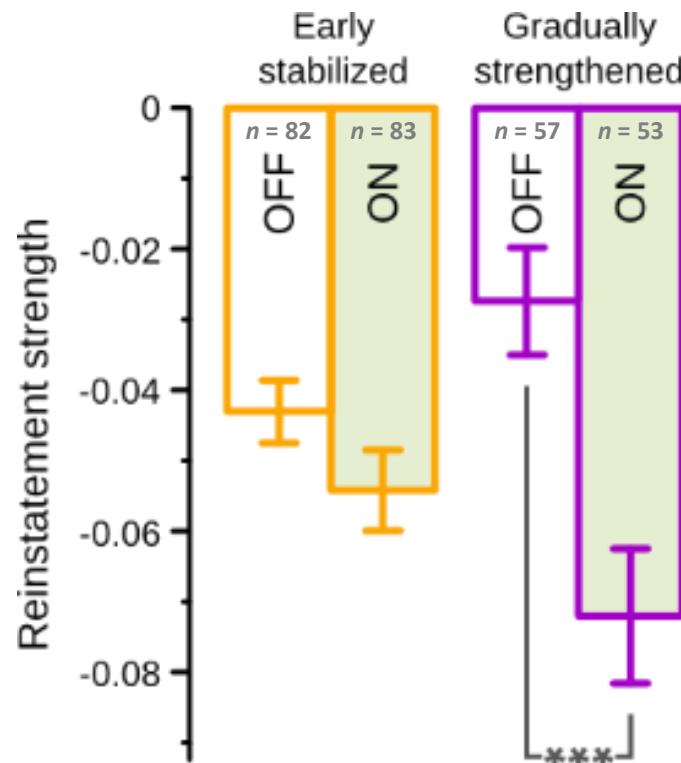
(based on 80 recording-blocks from 8 mice)

# Only stability of gradually strengthened patterns requires offline reactivation

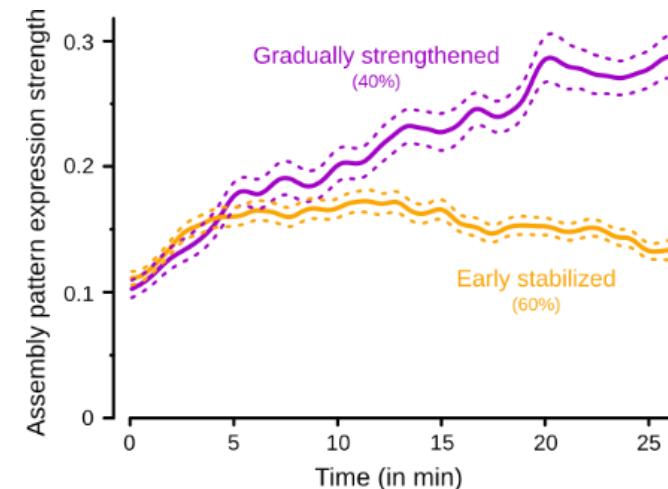


Gradually strengthened:  $n = 134$  assembly-patterns  
Early stabilized:  $n = 201$  assembly-patterns  
(based on 50 recording-blocks from 8 mice)

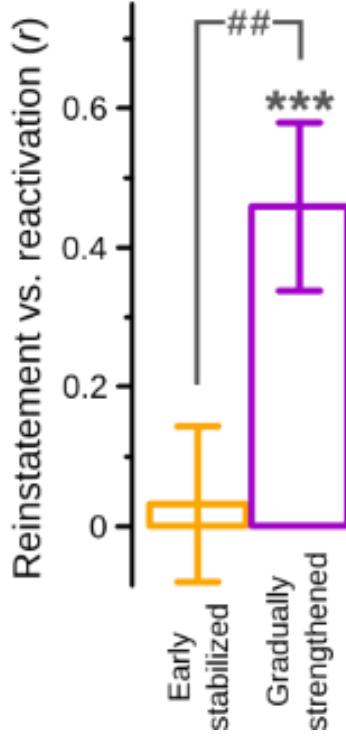
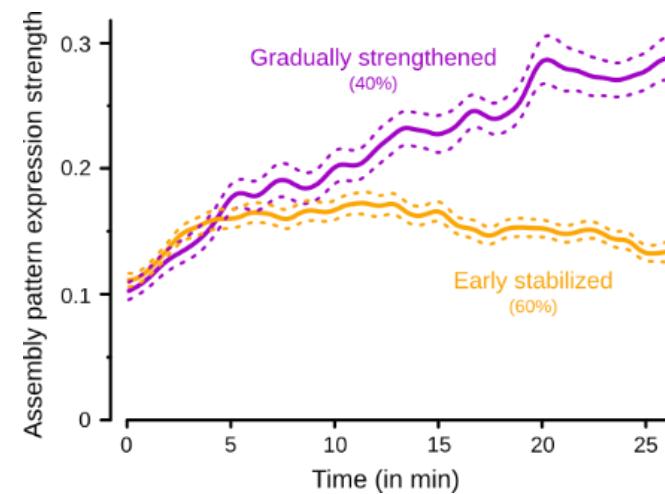
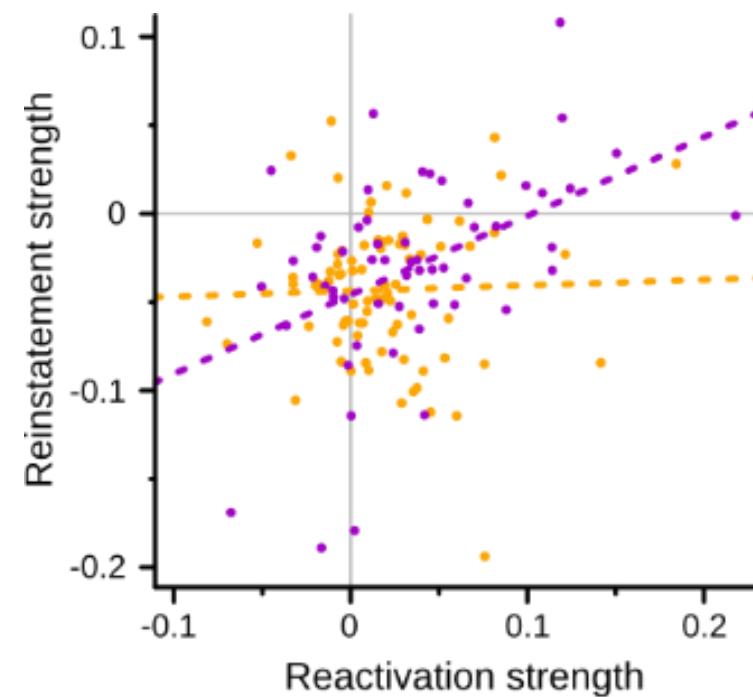
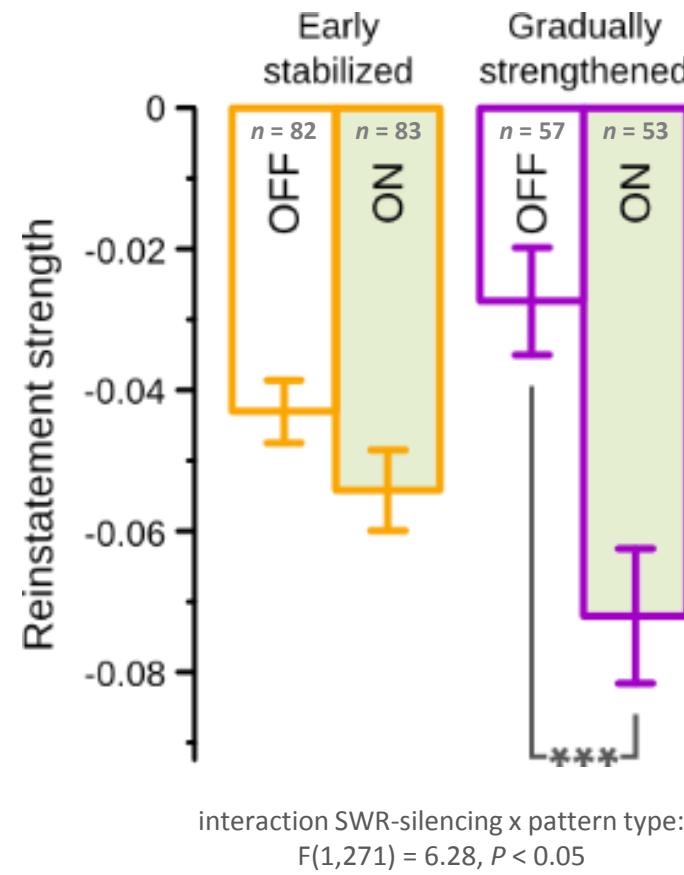
# Only stability of gradually strengthened patterns requires offline reactivation



interaction SWR-silencing x pattern type:  
 $F(1,271) = 6.28, P < 0.05$



# Only stability of gradually strengthened patterns requires offline reactivation



(based on 50 recording-blocks from 8 mice)

# One-sentence summary

The stability of “Hebbian-like” cell assembly patterns, which are gradually strengthened during their initial expression, depends on their offline reactivation.

## Acknowledgements

### Dupret lab

**David Dupret**

**Stephanie Trouche**

**Colin McNamara**

Natalia Campo-Urriza

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Alvaro Tejero-Cantero

Claire Bratley

Pavel Perestenko

Vadim Koren

Helen Barron

Alexander Morley

Mohamady El-Gaby

Stephen McHugh

### Visualizing tetrode tracks

Ben Micklem

Linda Katona

### Research support

Jane Janson

Liz Norman

Lisa Conyers

Katharine Whitworth

Kristina Wagner

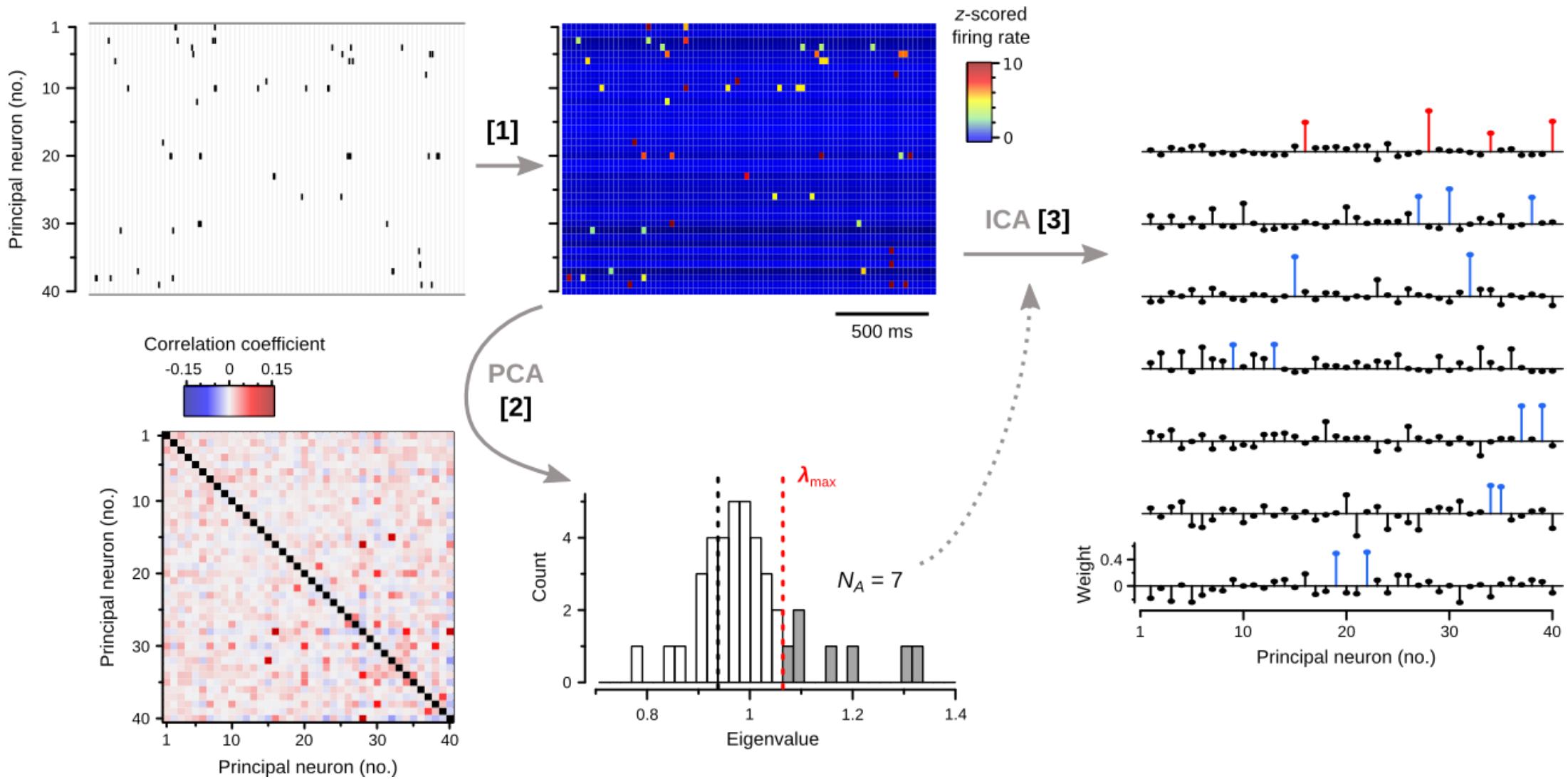
### Heidelberg

**Kevin Allen** (real-time SWR detection)

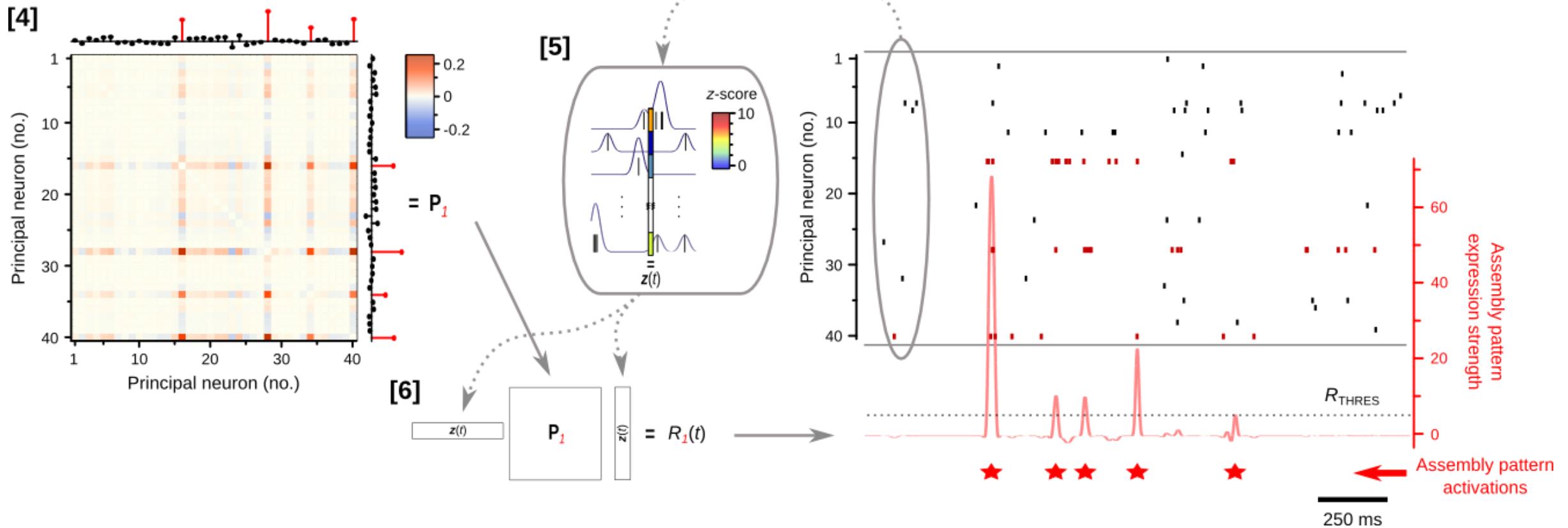




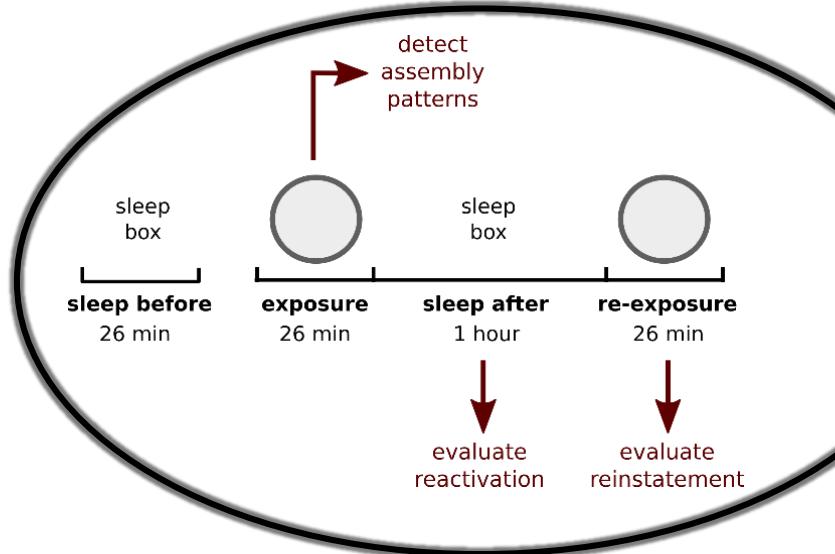
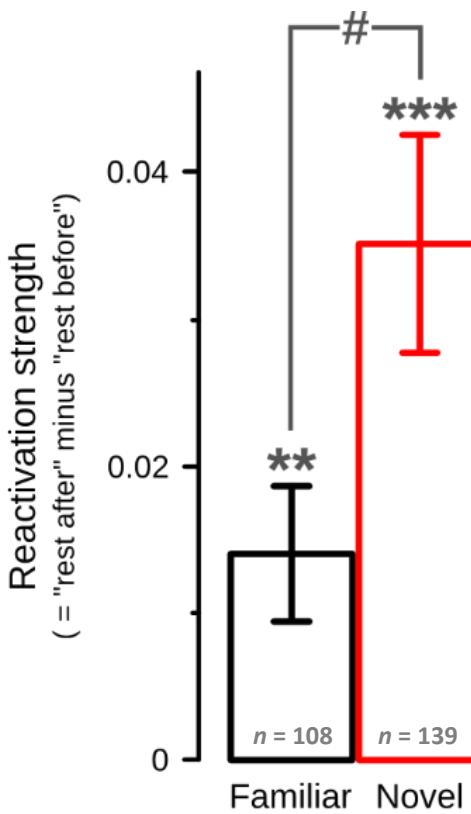
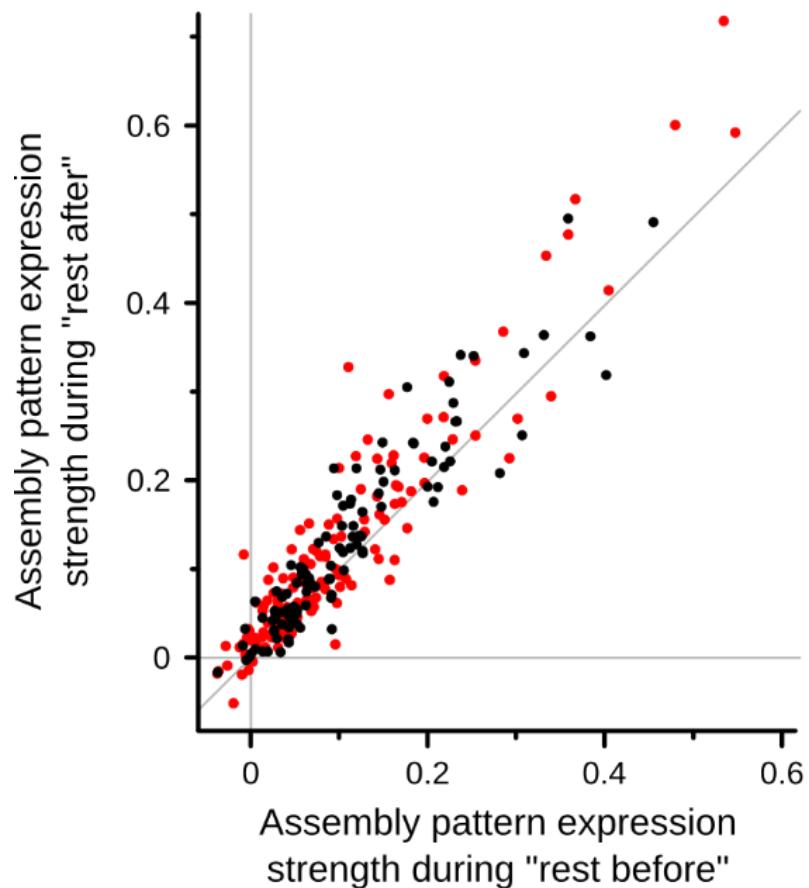
# Assembly pattern identification



# Assembly pattern tracking

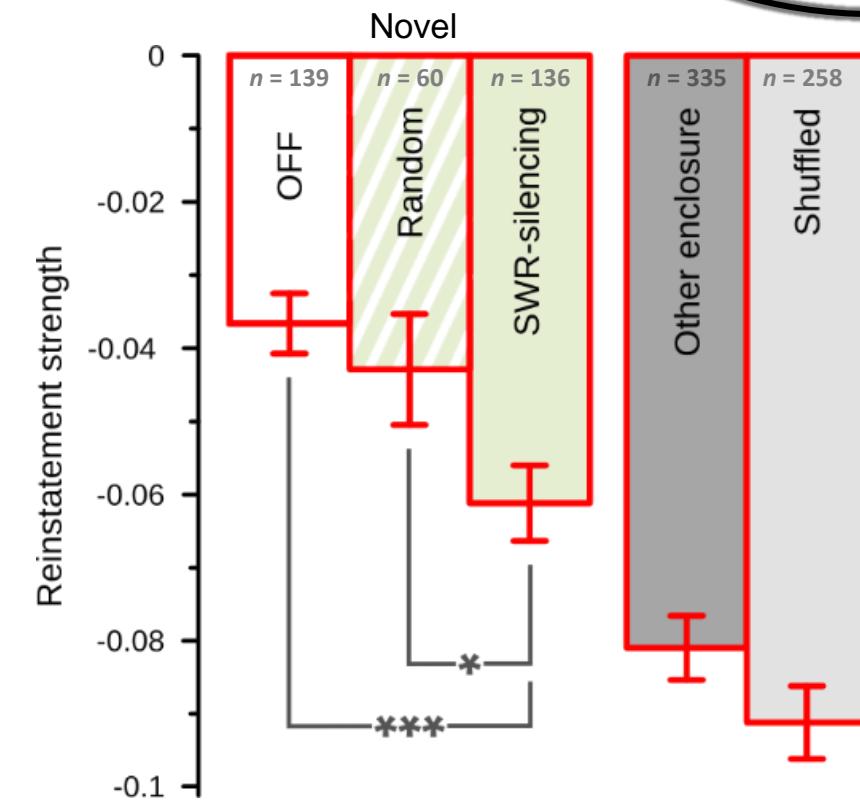
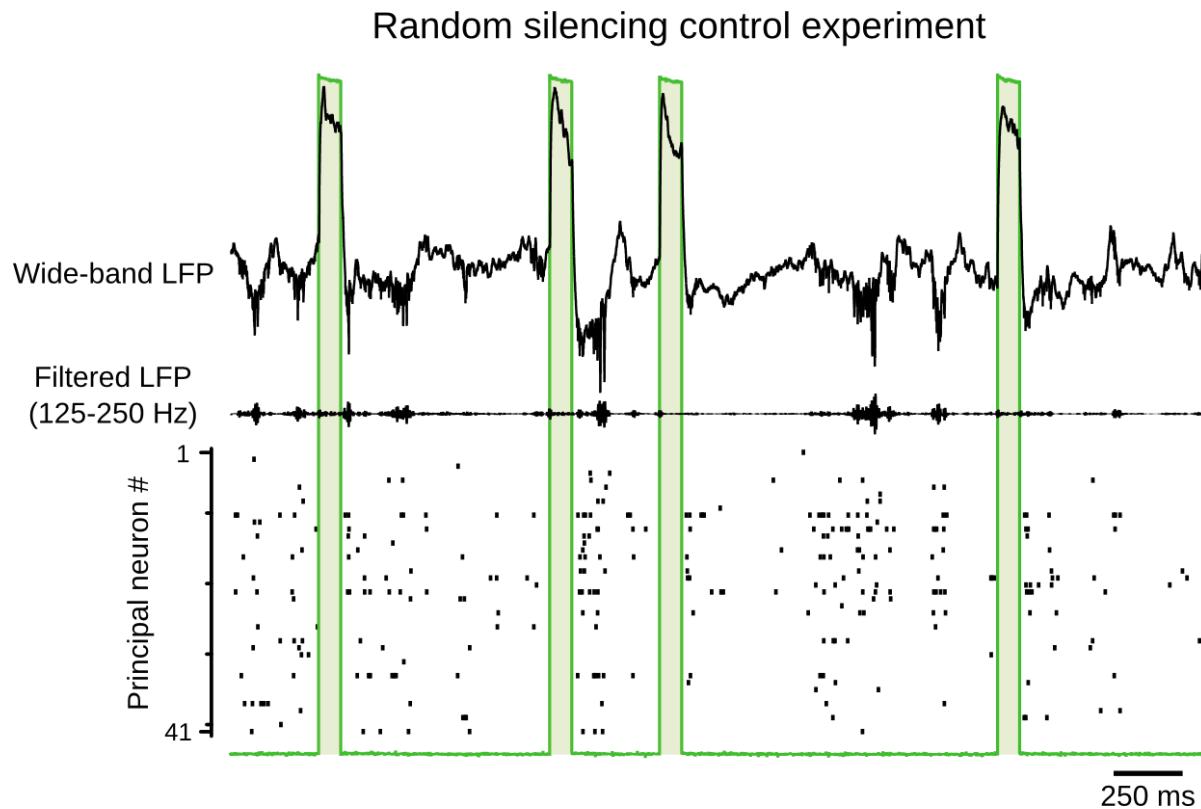
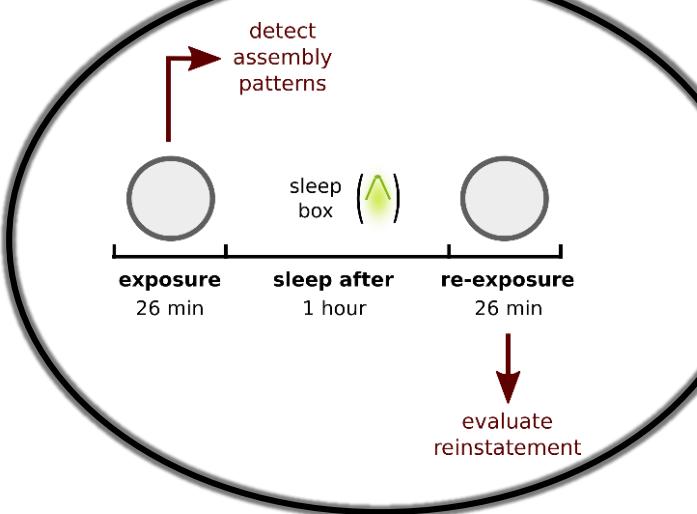


# Assembly pattern reactivation



(based on 43 recording-blocks from 8 mice)

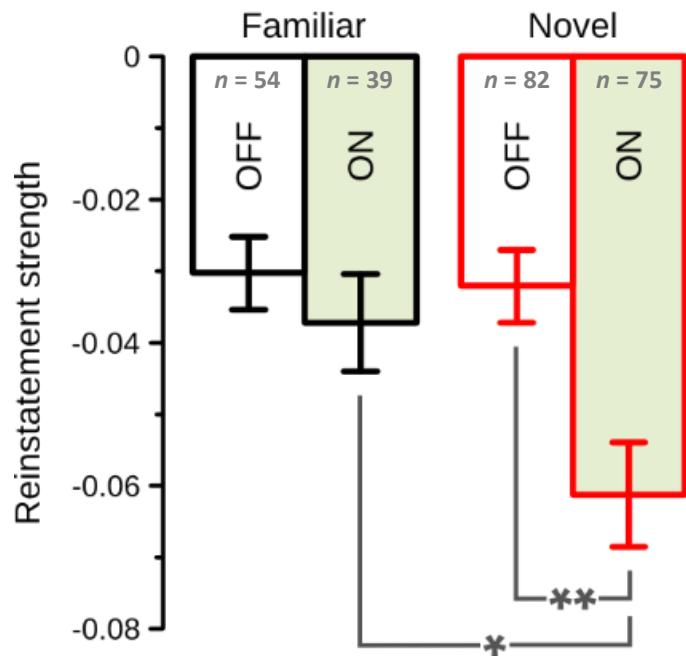
# Control experiment: random silencing



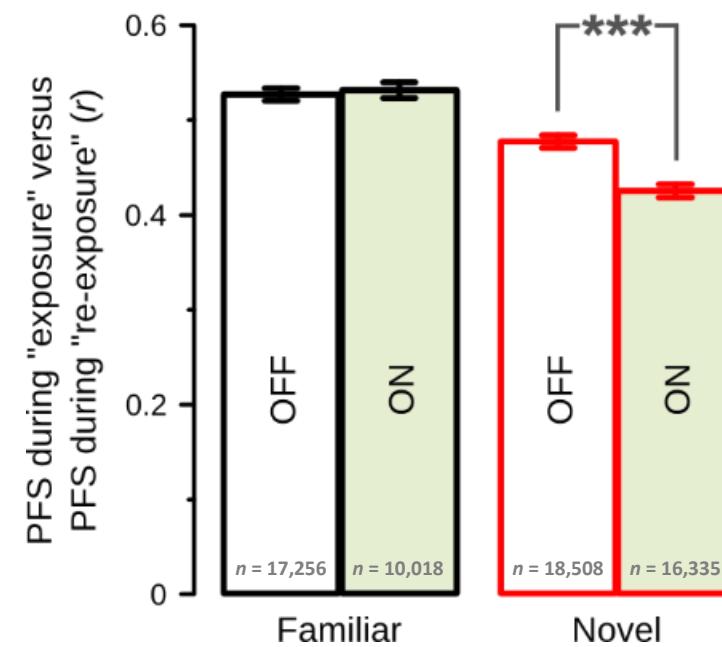
(based on 63 recording-blocks from 8 mice)

# Effect of SWR-silencing: control analyses

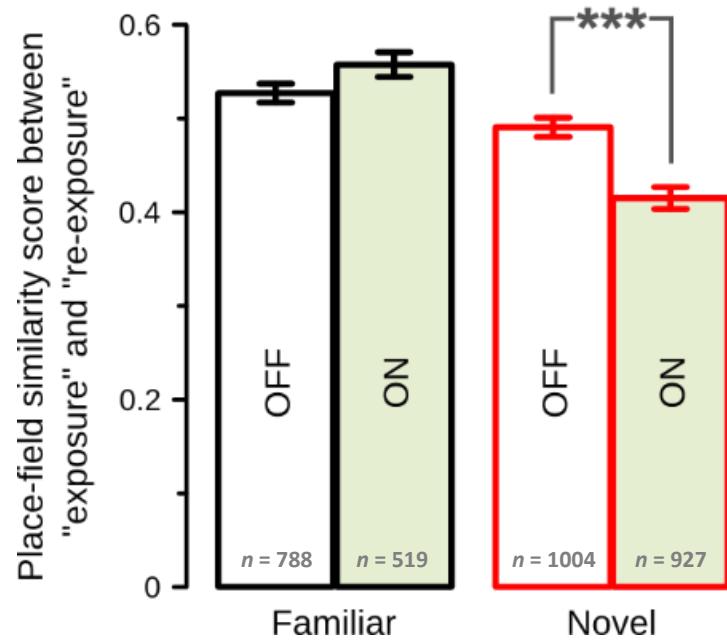
"Across-tetrodes only" analysis



Neuron-pair analysis



Single-neuron analysis



(based on 80 recording-blocks from 8 mice)